



# 九十九學年度 微生物學教材

細菌遺傳學、細菌基因體學

(11-16-2010, 11-17-2010)

# Chapter 14

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Bacterial Genetics: 14.4  
Creating genetic variability

11-16-2010

<http://life.nctu.edu.tw/~hlpeng/>

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# 課程綱要與評分標準

11/16, 11/17	Bacterial genetics and microbial genomics
11/23, 11/24	Microbial taxonomy, Archaea
11/30	Fungi
<u>12/7</u>	<u>Exam I- 20%</u>
12/8	Bacteria (I)
12/14, 12/15	Bacteria (II)
12/21, 12/22	Viruses (I)
12/28, 12/29	Viruses (II), viroids, virusoids, and prions
1/4, 1/5	Microbial diseases and their control
<u>1/11</u>	<u>Exam II- 20%</u>

出席率      10.5% (21 h)

# NDM-1 (*New Delhi Metallo-beta-lactamase*)

- In December 2009, a carbapenem-resistant *K. pneumoniae* infection acquired in New Delhi, India
    - Resistant to multiple antibiotics
    - Also reported in *E. coli* and *Acinetobacter baumannii*
- The Lancet infectious disease 2010*



NDM-1 Kp01

超級細菌



# Horizontal (Lateral) Gene Transfer (HGT) in bacteria

- Haploid genome
- Exogenote (Donor)
- Endogenote (Recipient)
- Merozygote
  - Transconjugants
  - Transformants
  - Transductants

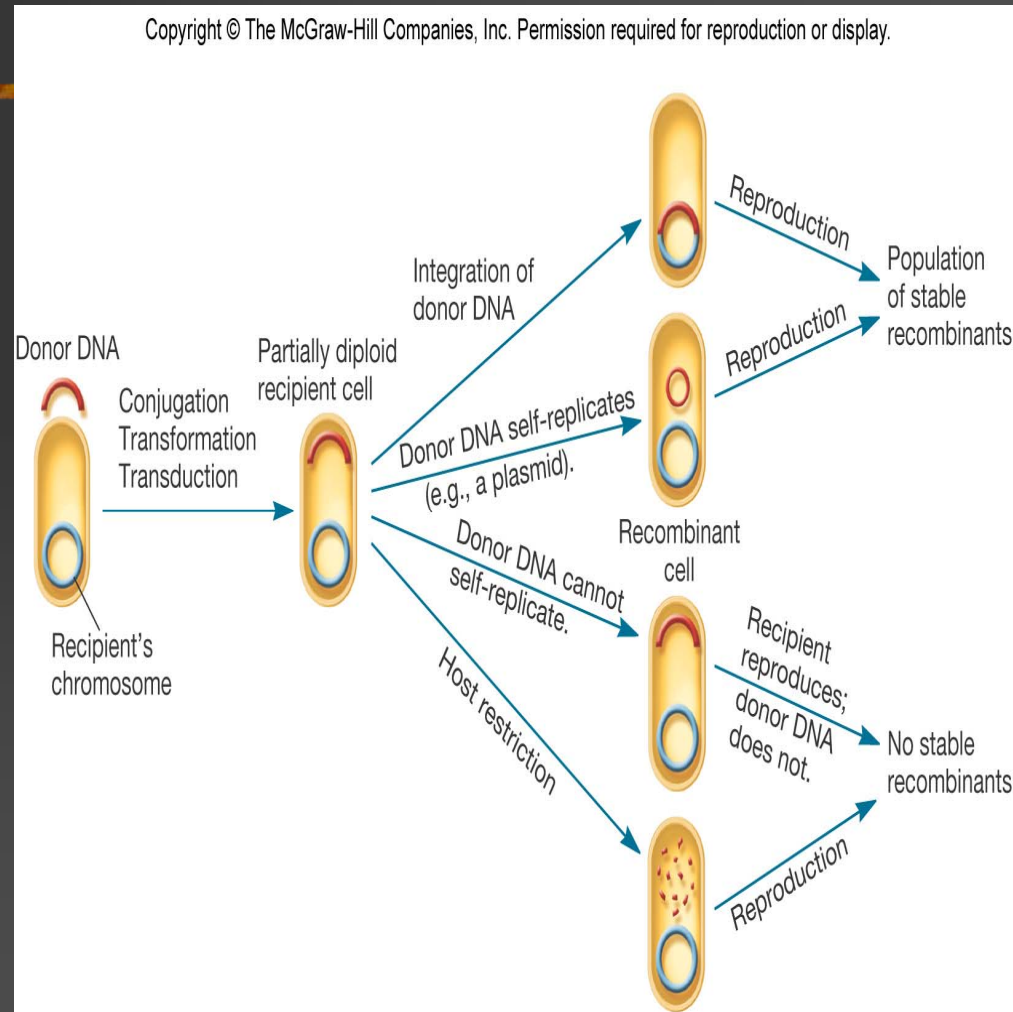
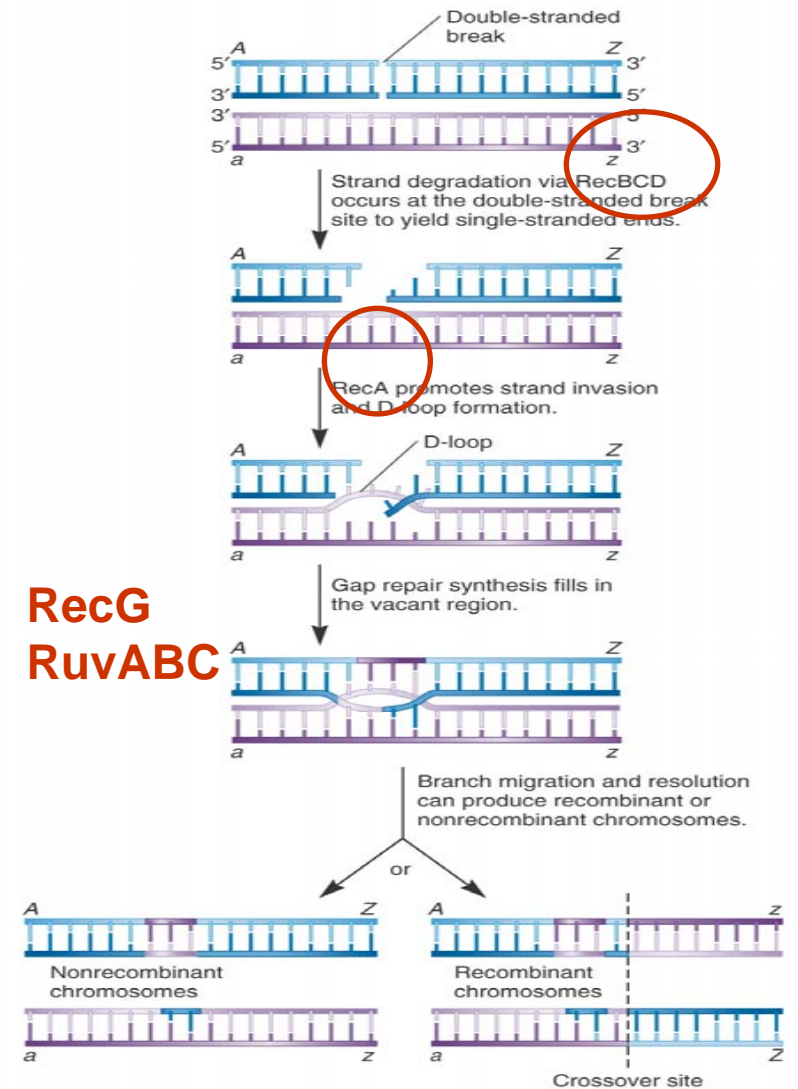


Figure 14.11

# Recombination at the molecular level

- Homologous recombination (Reciprocal recombination)
  - **Double-Strand Break Model**
    - results from DNA strand breakage and reunion, leading to crossing-over

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# Nonreciprocal Homologous Recombination

- incorporation of single strand of DNA into chromosome, forming a stretch of heteroduplex DNA
- proposed to occur during bacterial transformation

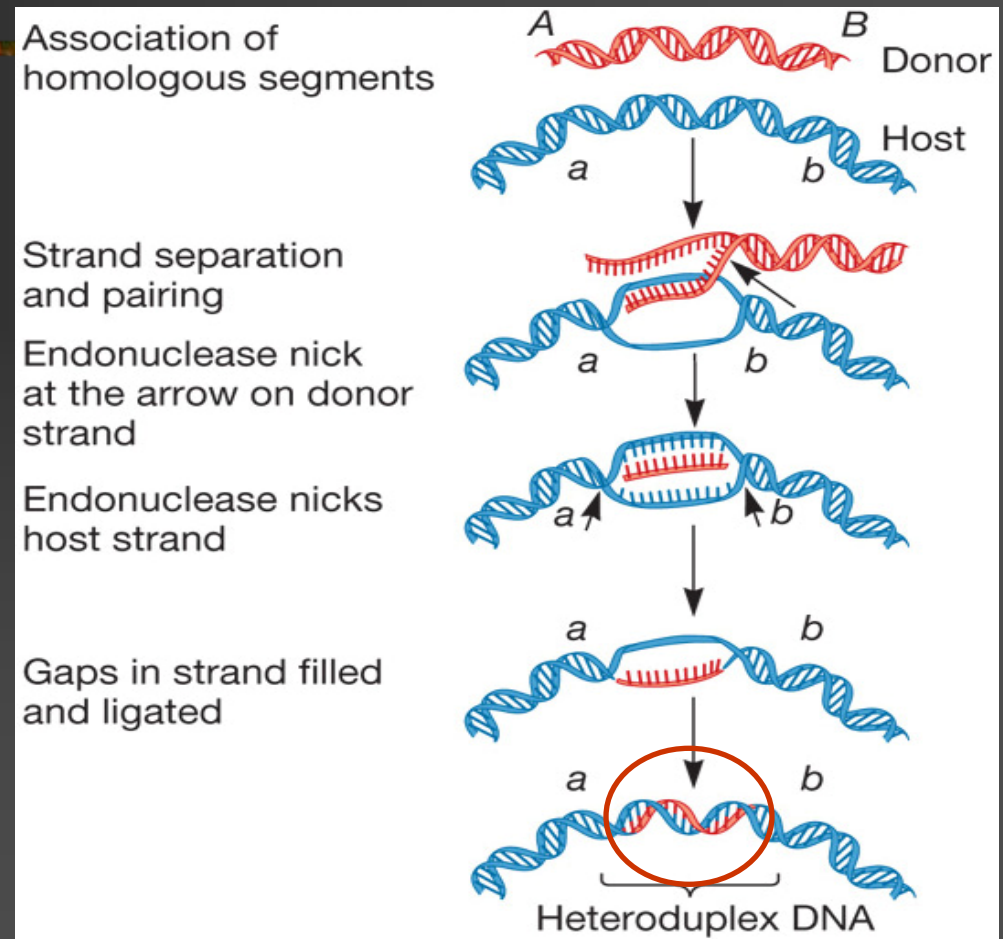


Figure 14.13

# Site-specific recombination

- insertion of viral genome into host chromosomes
  - insertion of **nonhomologous DNA**
    - Only a small region of homology is required
- Transposition carried out by **transposable elements**
  - Simple transposition
  - Replicative transposition

# Transposable Elements

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- **transposition**

- the movement of pieces of DNA around the genome

- **transposable elements (transposons)**

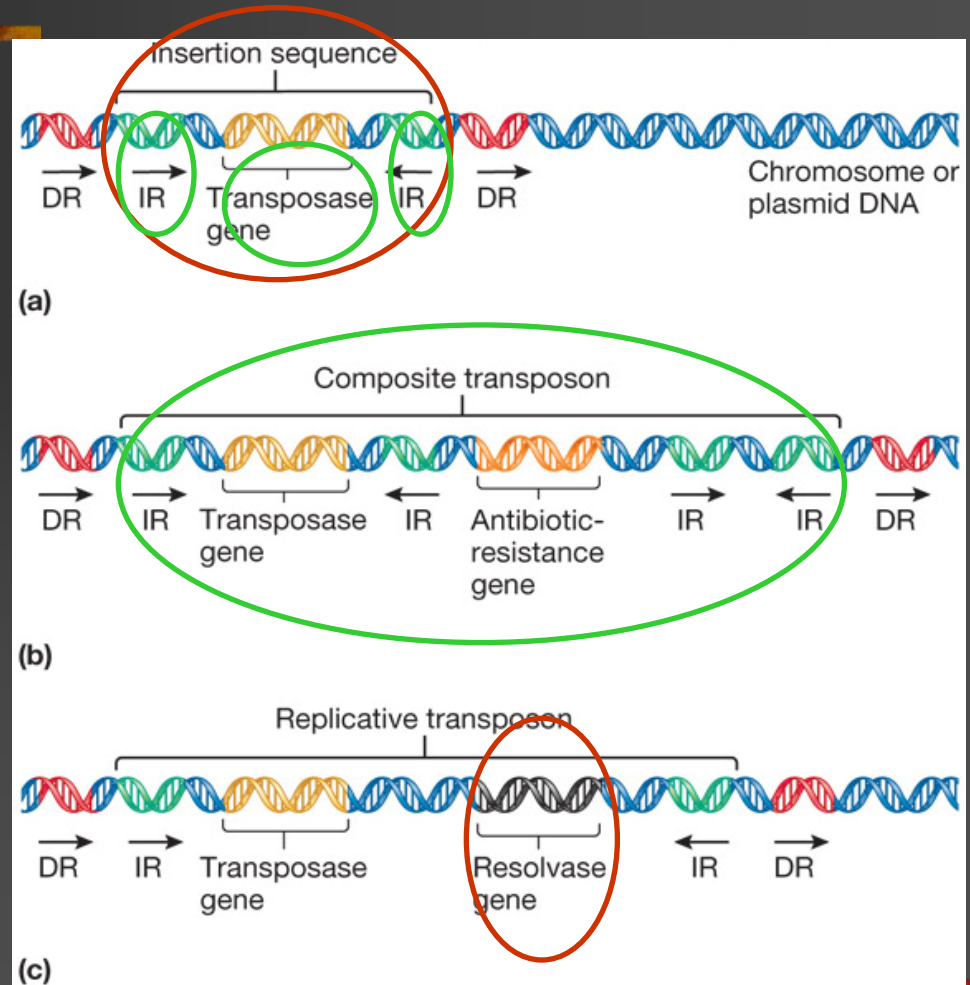
- segments of DNA that carry genes for transposition

- widespread in bacteria, eukaryotes and archaea

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# Types of transposable elements

- IS element
  - 2 IR + transposase gene
- Transposons
  - 2 IS elements
  - additional genes
- Replicative transposons
  - Resolvase gene



# Simple Transposition- cut and paste

- Generation of **direct repeats** in host DNA flanking a transposon

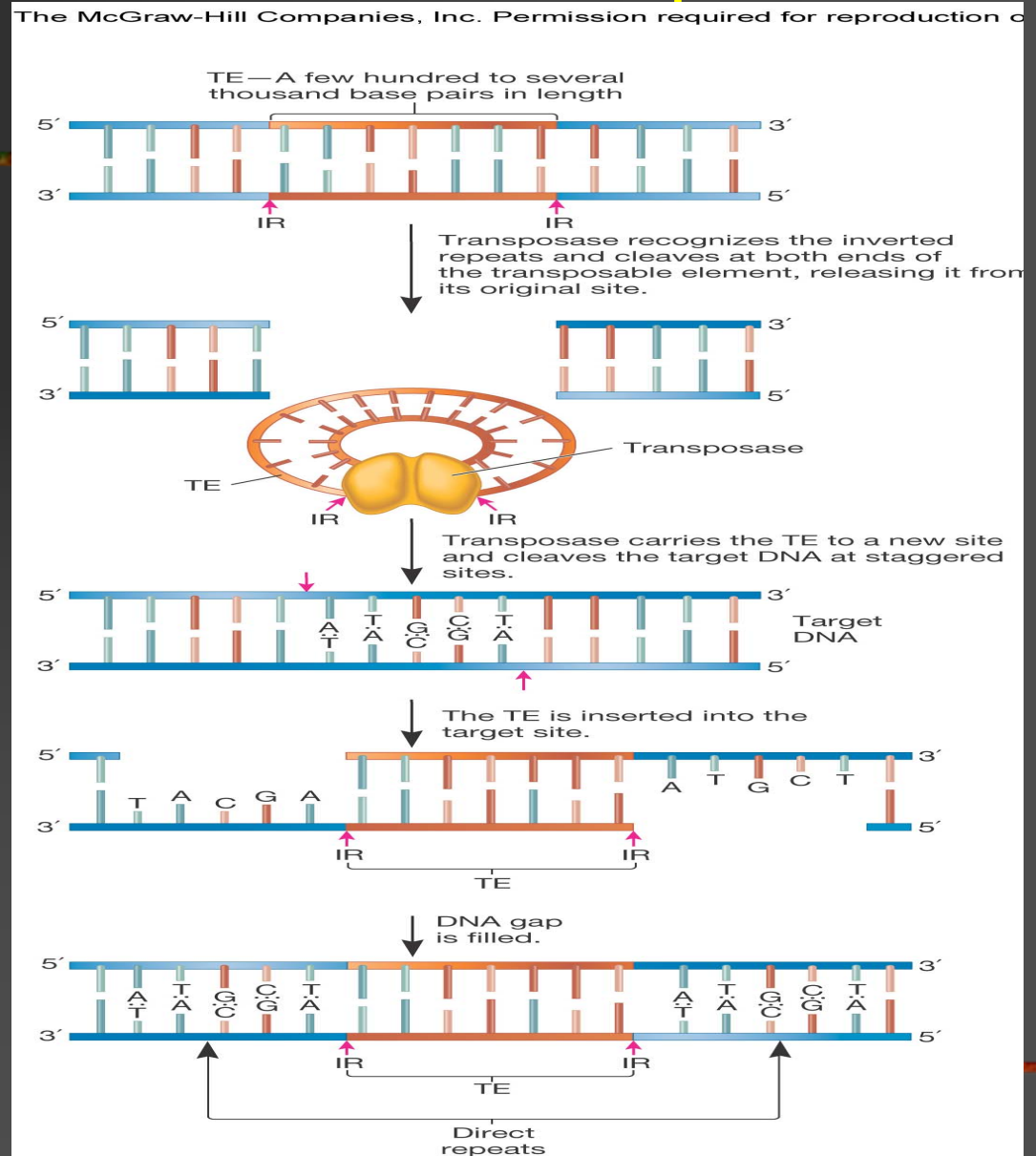


Figure 13.20



# Replicative Transposition

- insertion generates direct repeats of flanking host DNA
- usually transposon replicated, remaining in original site, while duplicate inserts at another site
- Tn3

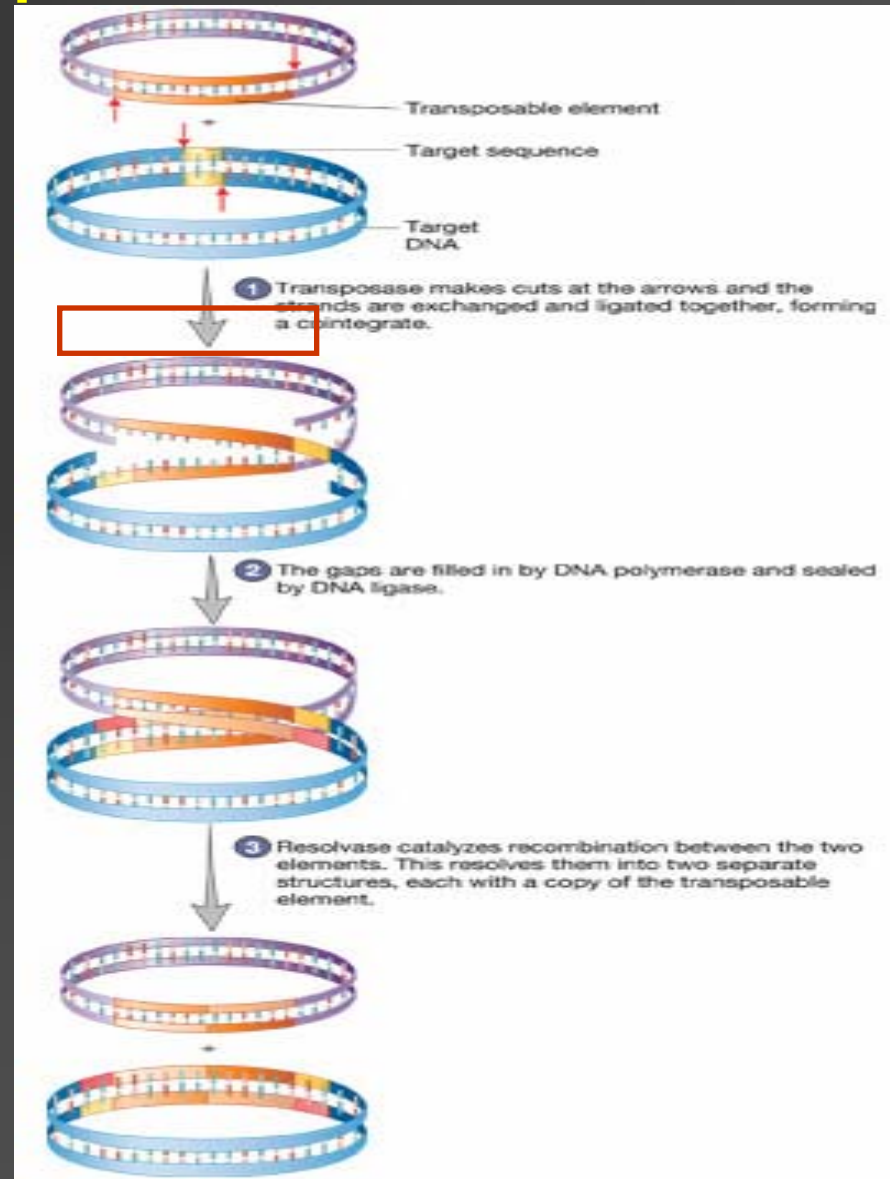


Figure 14.16

# Effects of transposition

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- mutation in coding region
  - arrest of translation or transcription
    - interruption of **promoter** regions...
    - interruption of **ribosomal binding sites**...
  - activation of genes
    - provide strong **promoter** sequences
  - generation of new plasmids
    - **Multi-drug** resistance plasmids
-

# R Plasmid results from Tn3 Transposition

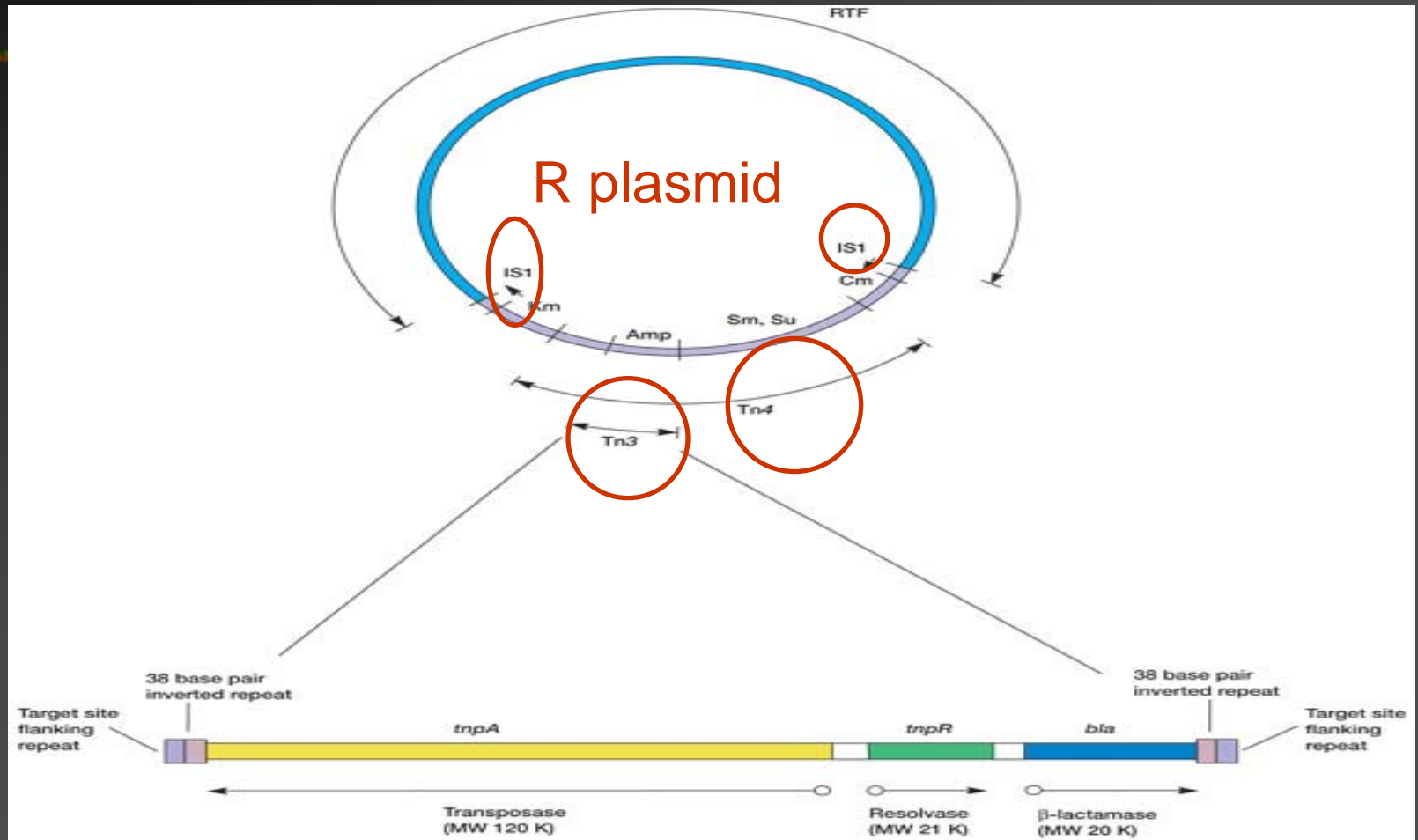


Figure 14.17

# Plasmids

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- small, autonomously replicating DNA molecules that can exist independently or, as episomes, integrate reversibly into the host chromosome
  - conjugative plasmids such as the F plasmid can transfer copies of themselves to other bacteria during conjugation
  - R plasmid, virulence plasmid, metabolic plasmid...

# Bacterial conjugation

- The transfer of genes between bacteria that depends on direct cell to cell contact
  - mediated by the sex pilus encoded by **F plasmid**

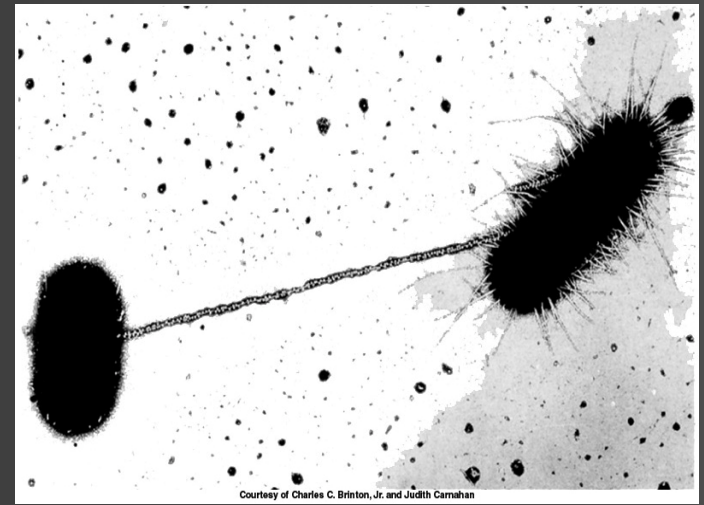


Fig. 14.19

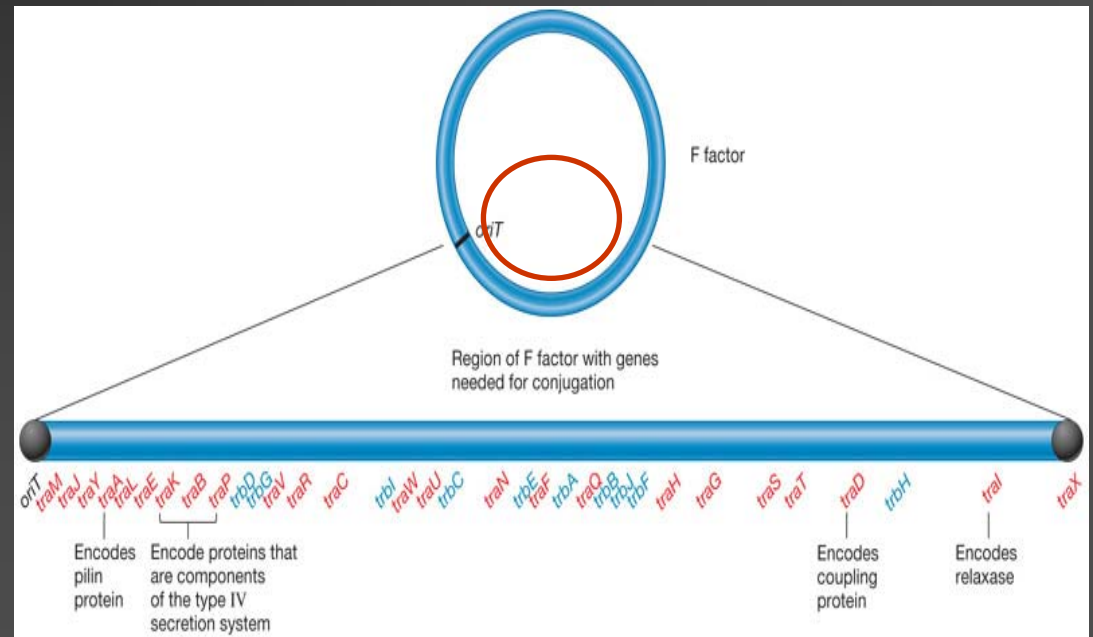


Fig. 14.18

# F plasmid integration

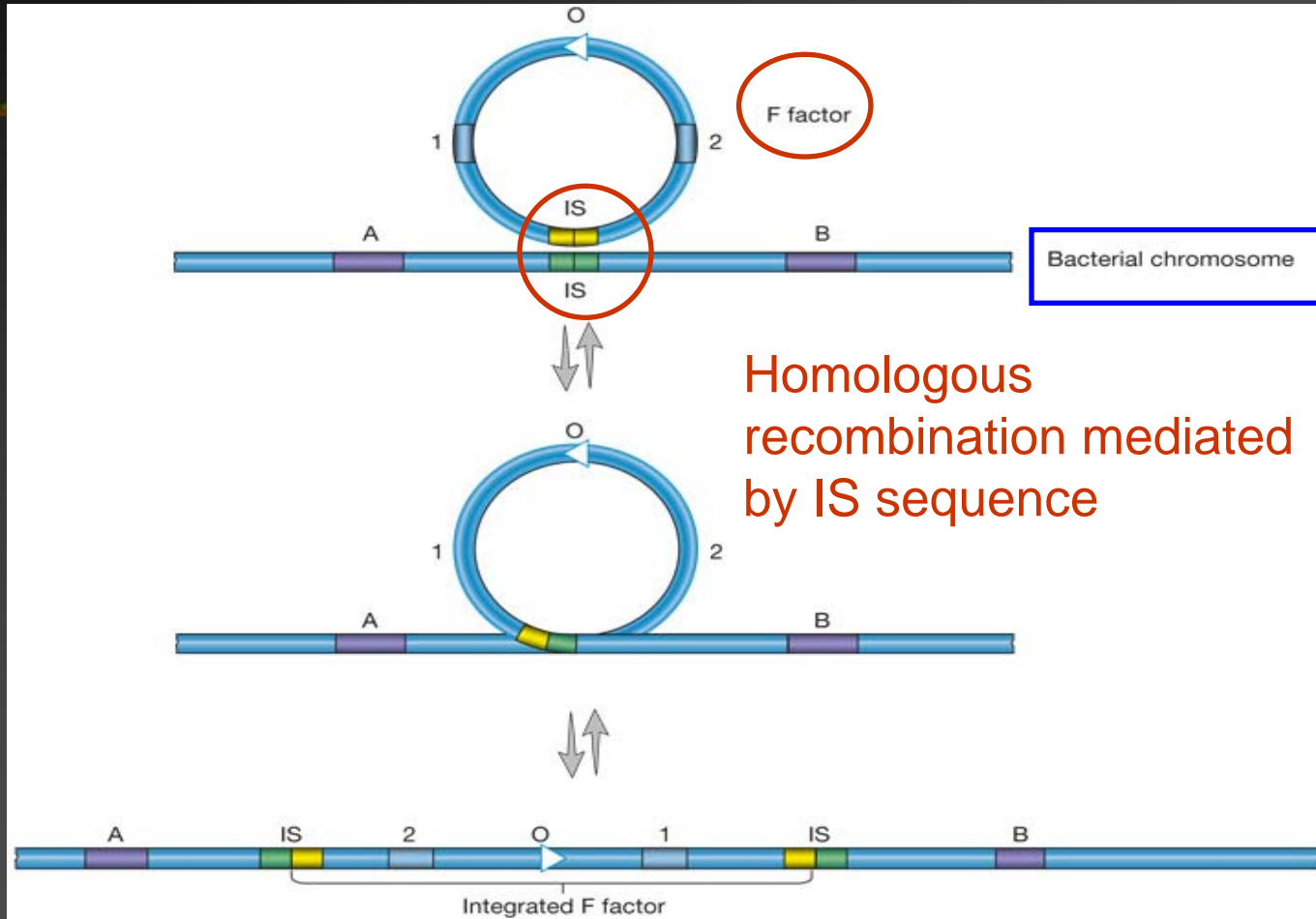
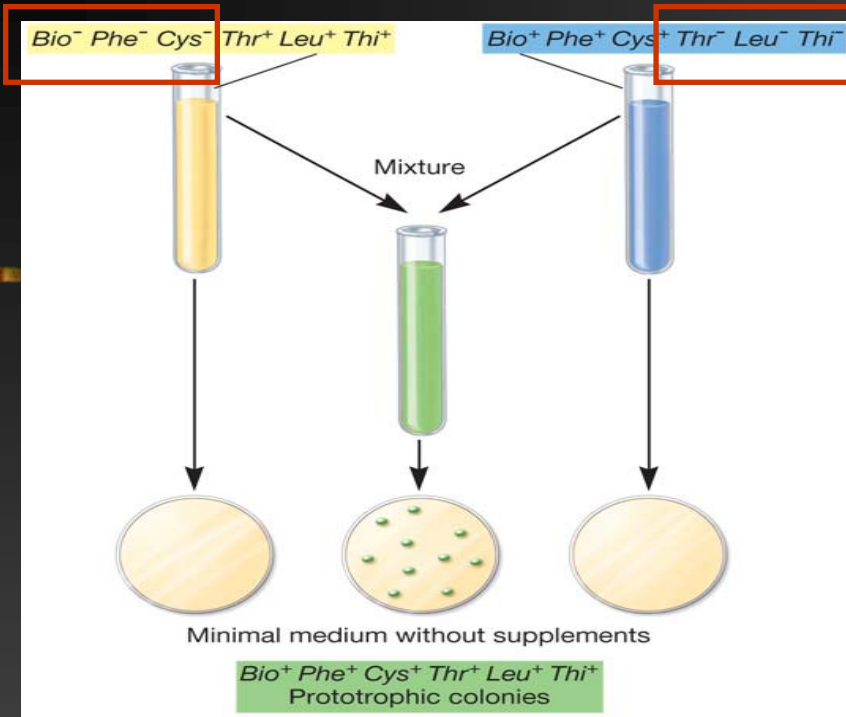
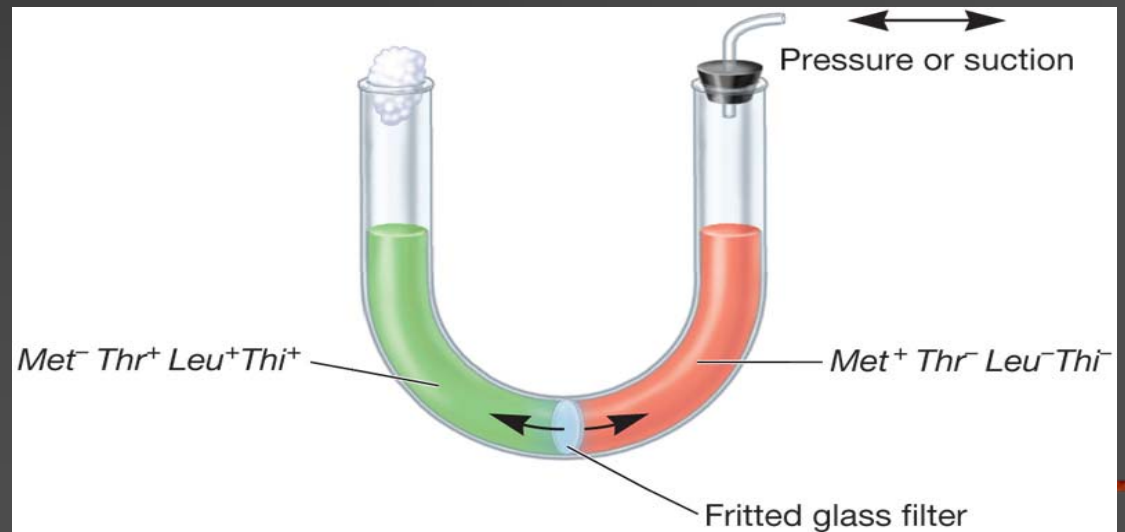


Figure 14.20



- transfer of DNA between cells discovered by Lederberg and Tatum (1946)

## The U-tube experiment



- cell-to-cell contact is necessary for the DNA transfer, which demonstrated by Bernard Davis (1950)



# F-mediated conjugation

- $F^+ \rightarrow F^-$ 
  - A polar gene transfer
  - replicated by **rolling-circle** and the duplicate is transferred
  - recipients usually become  $F^+$
  - donor remains  $F^+$

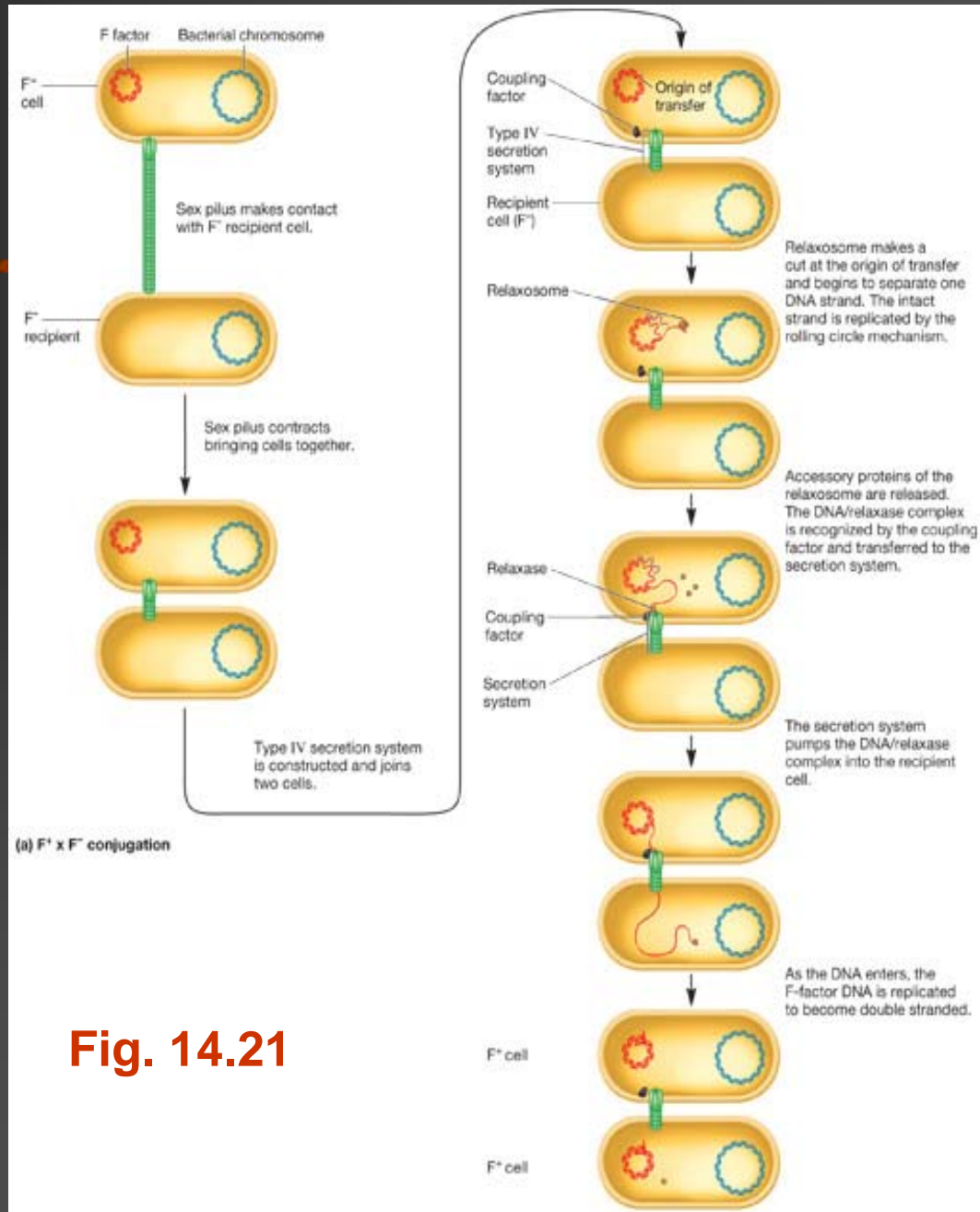


Fig. 14.21

# The type IV secretion system

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- Tra proteins
  - TraA pilin

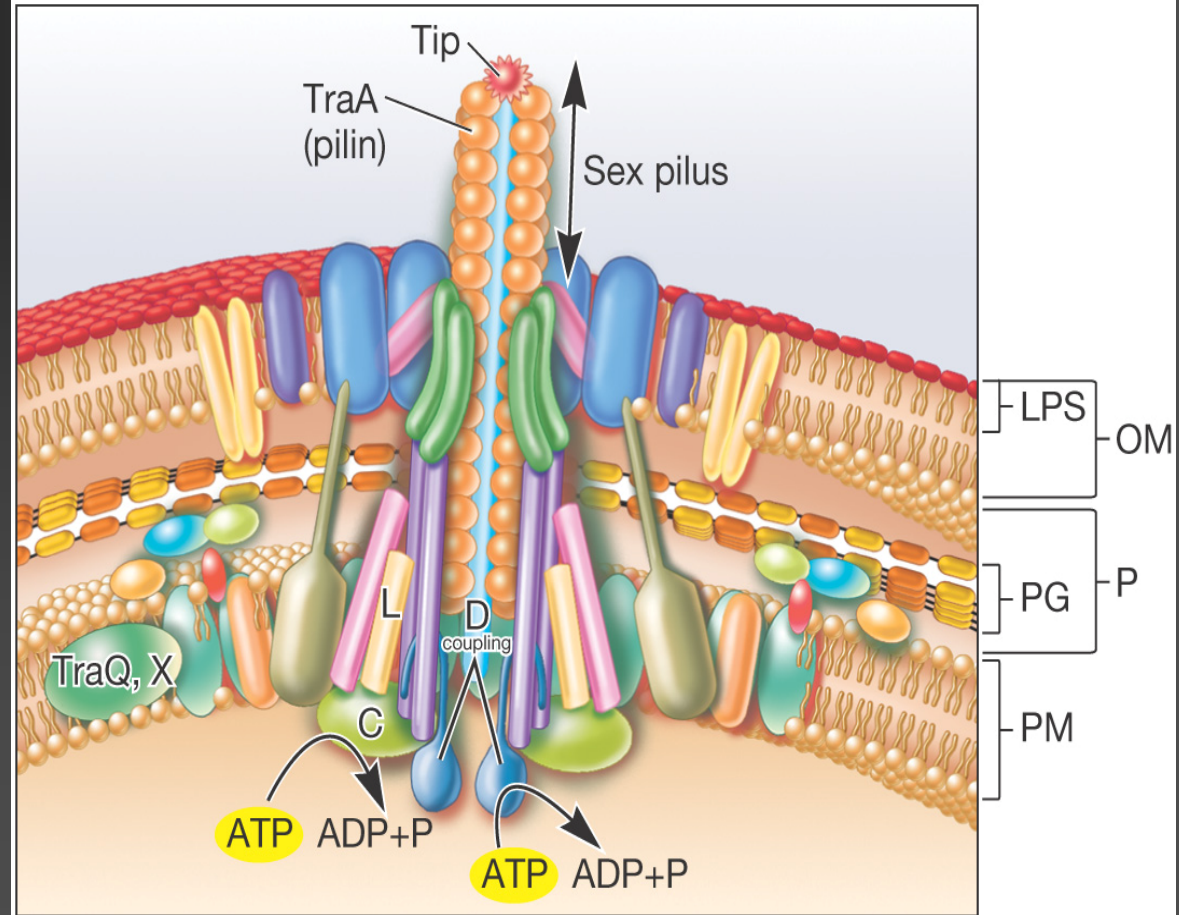


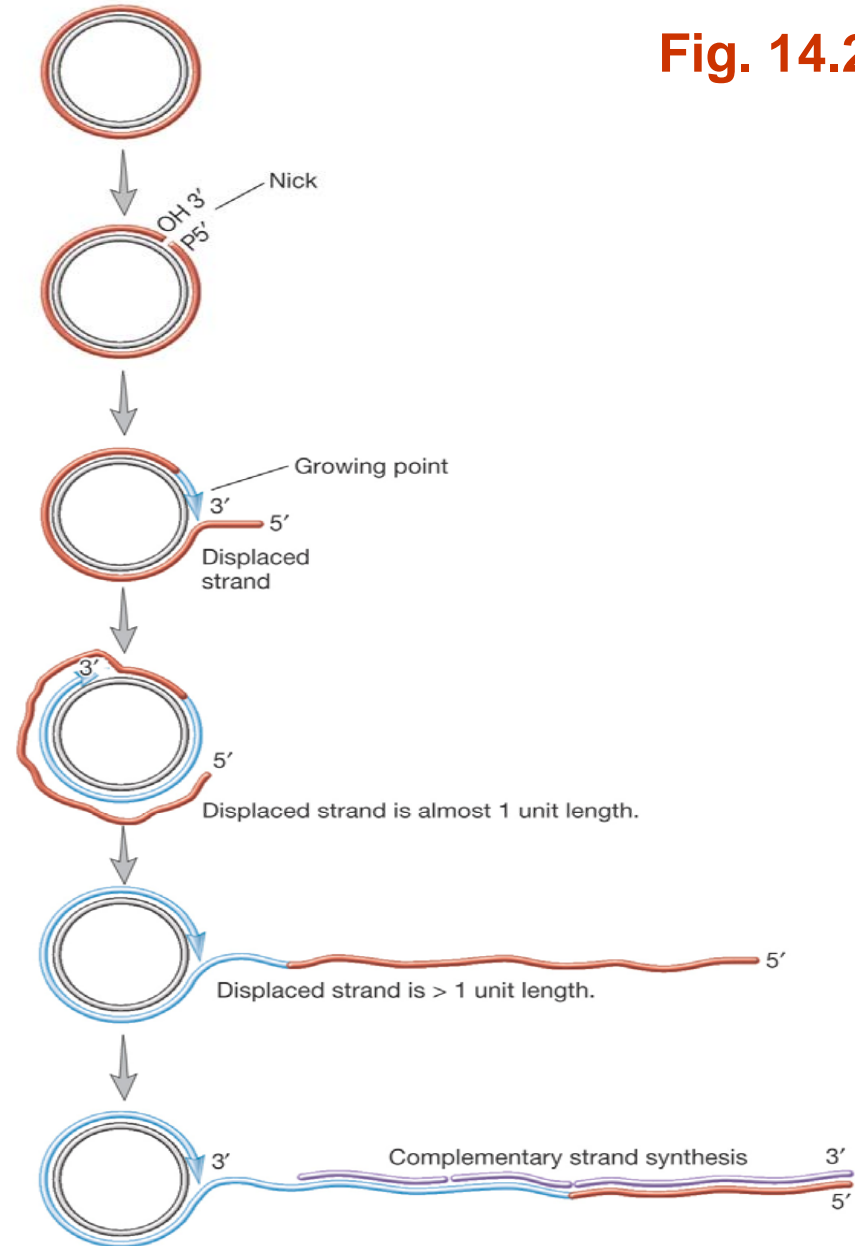
Fig. 14.22

# Rolling-circle replication

- A 1S tail is generated → 2S by synthesis of a complimentary strand

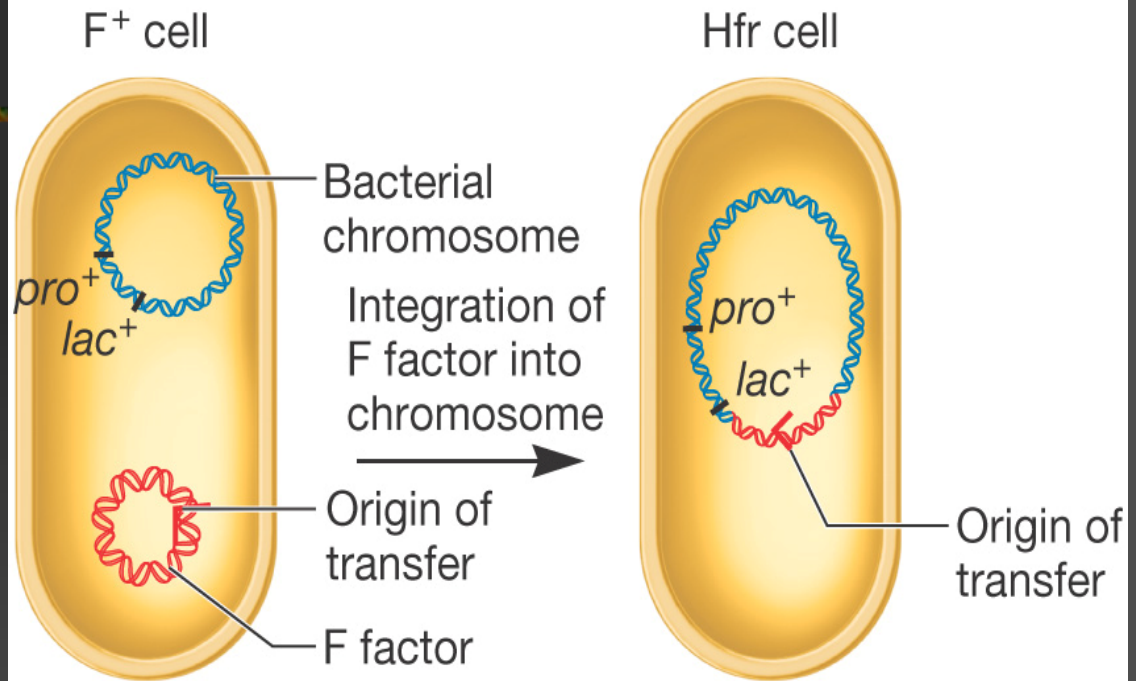
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Fig. 14.23



# Hfr cells

- Integration of F to chromosome to generate Hfr cell
- *high frequency* of recombinants



(a) Insertion of F factor into chromosome

# HFr → F<sup>-</sup> conjugation

- a complete copy of the F factor is usually not transferred

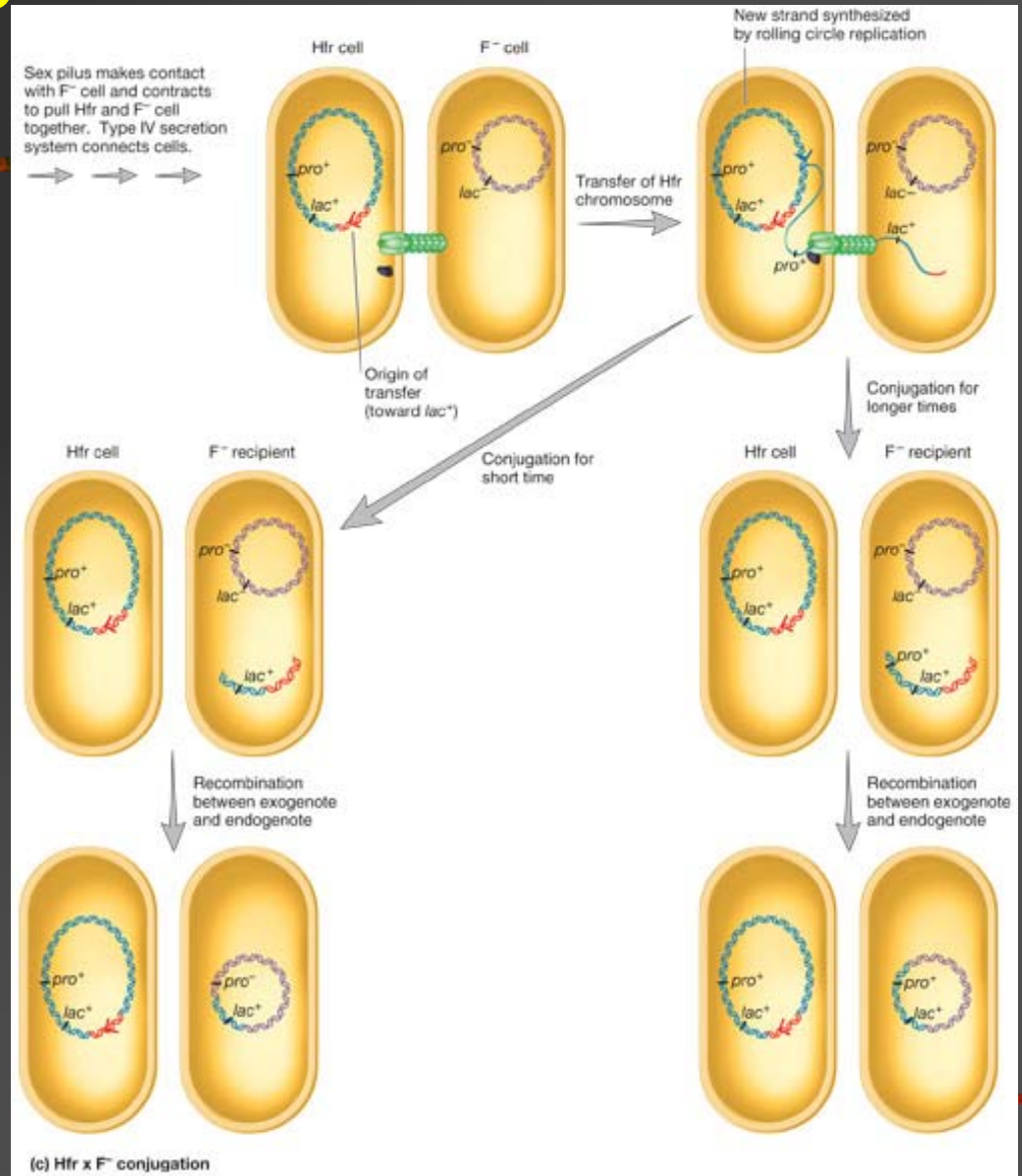


Fig. 14.24 (b)



# F' cells

- An error excision caused the **A** gene of Hfr cell is picked up by the F factor (**red**)

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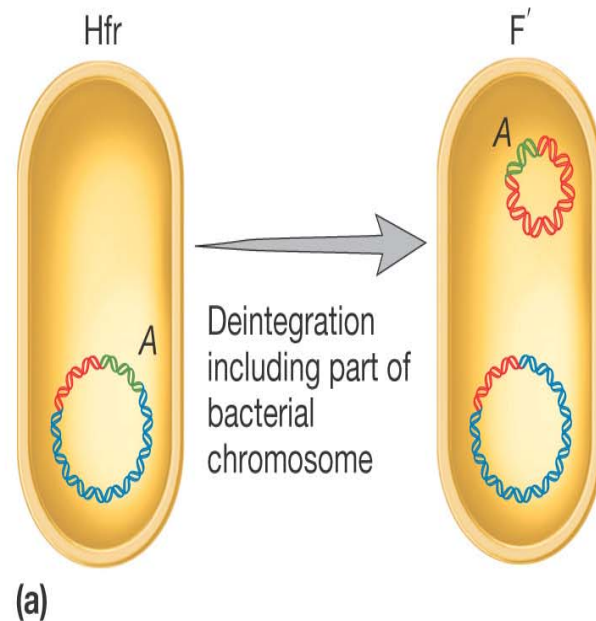


Fig. 14.25(a)

# F' conjugation

## F' x F- Mating

- formed by incorrect excision from chromosome
- some of the F factor is left behind in the host chromosome
- some host genes have been removed along with some of the F factor
  - these genes can be transferred to a second host cell by conjugation

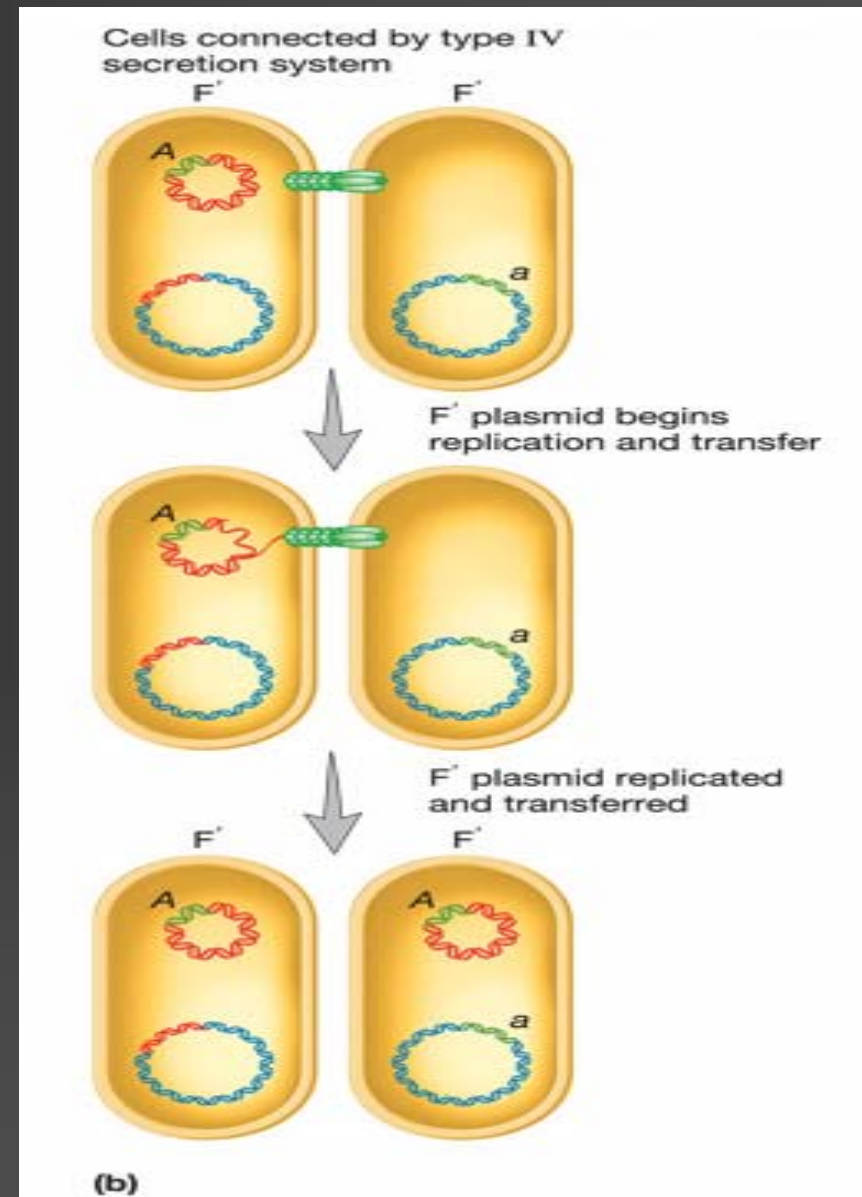
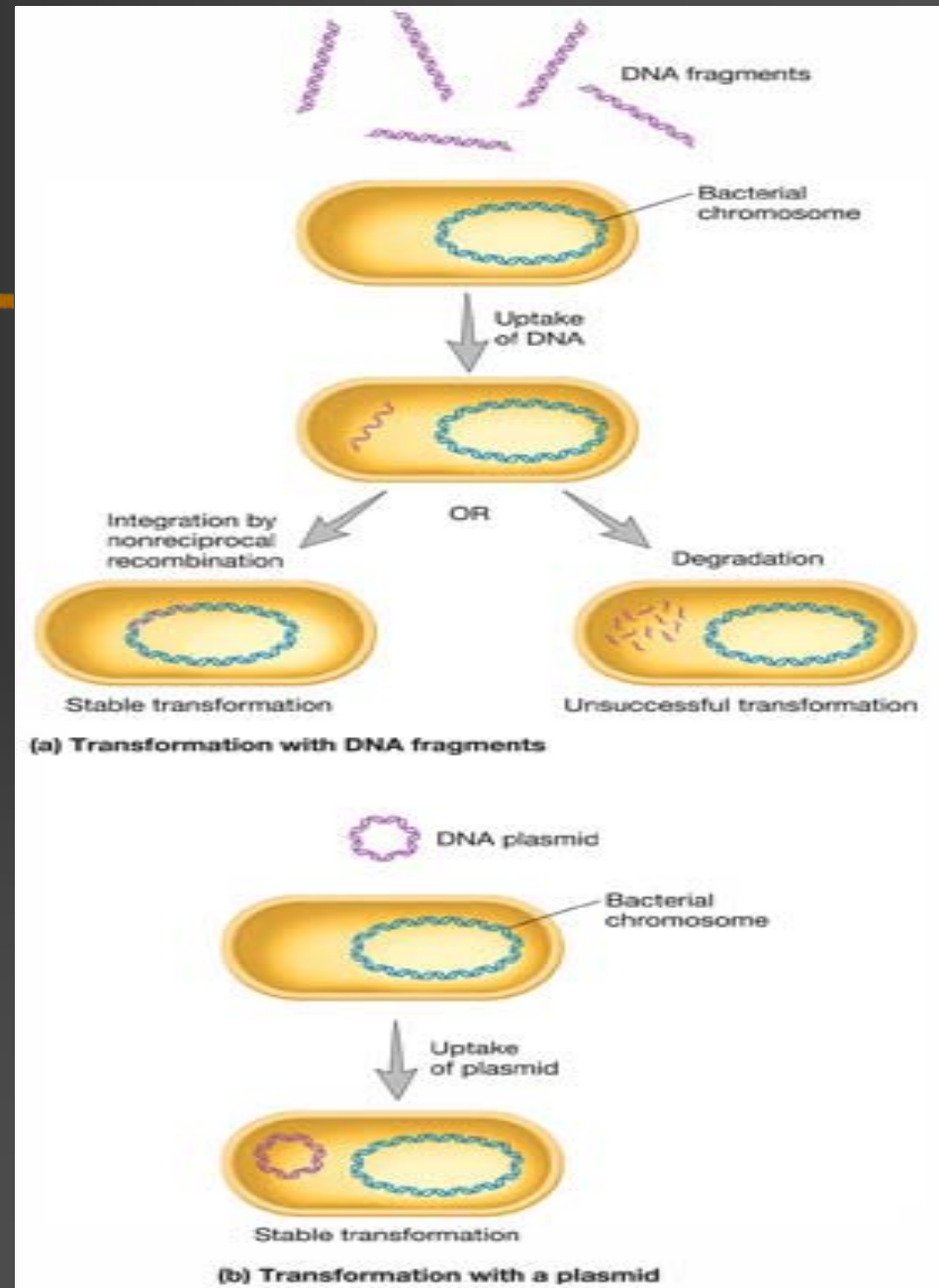


Fig 14.25 (b)



# DNA transformation

- uptake of DNA fragments and incorporation into recipient
  - competent cell
- uptake of a plasmid



# Transformation in *Streptococcus pneumoniae*

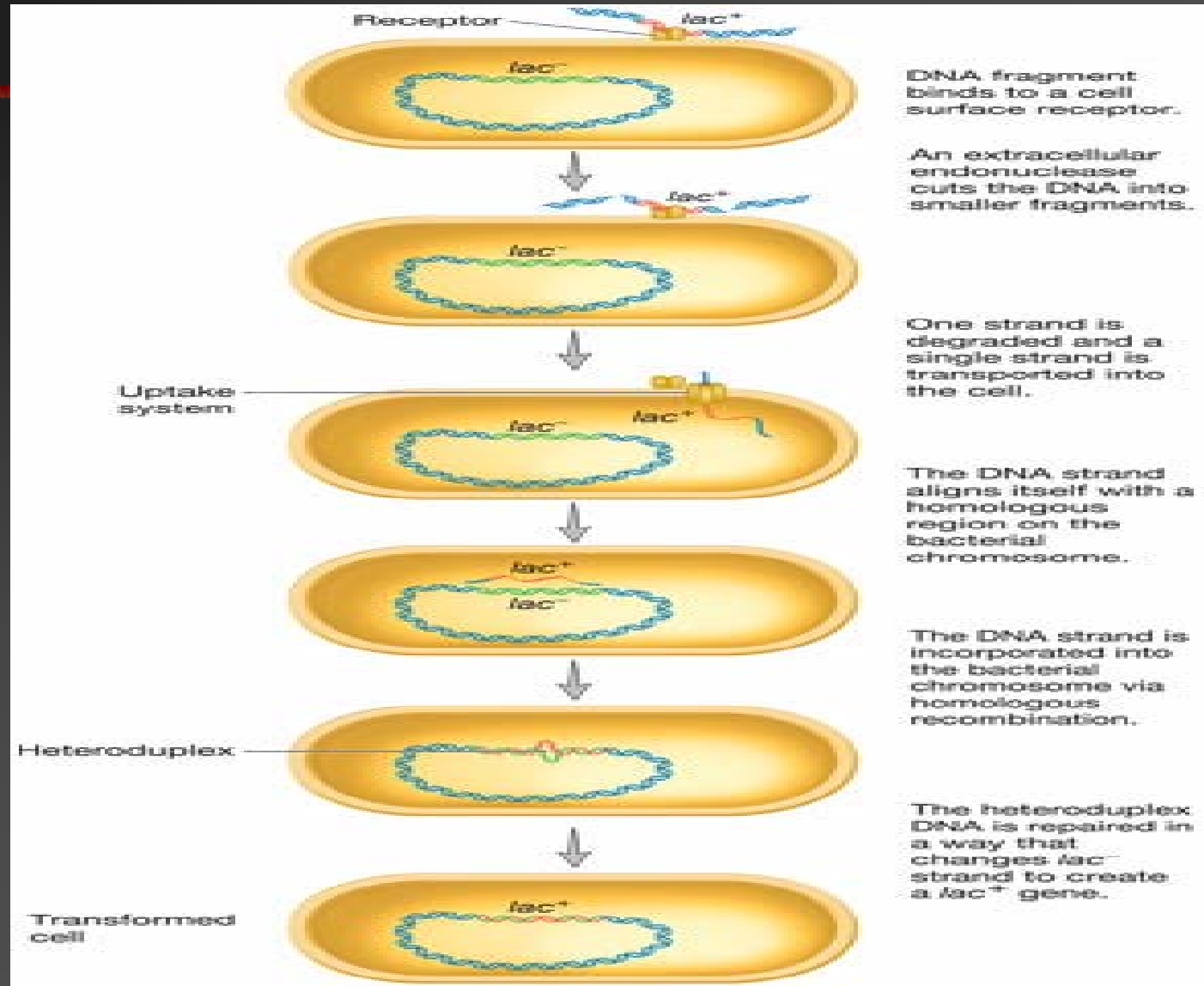
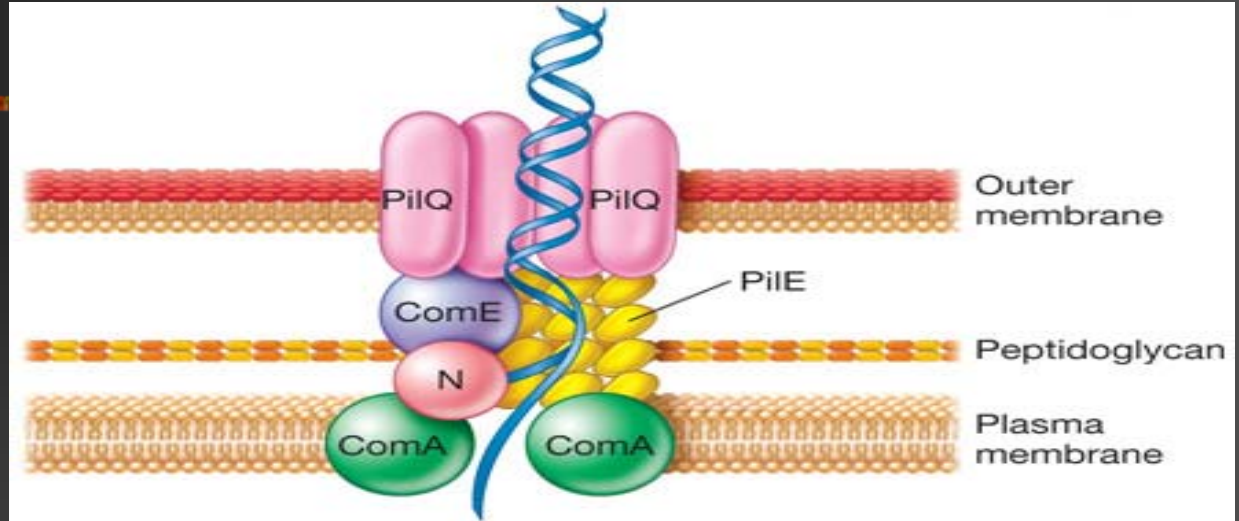


Fig 14.27

# DNA Uptake Systems

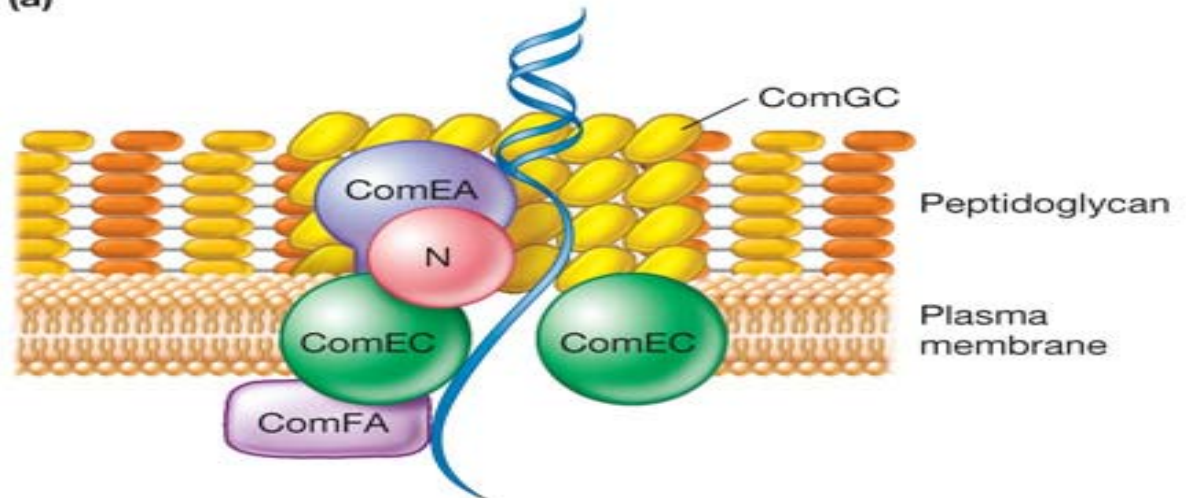
Natural competence

*G(-) Neisseria sp.*



(a)

*G(+) Bacillus sp.*



(b)

Fig 14.28

# Transduction- transfer of genes by phages

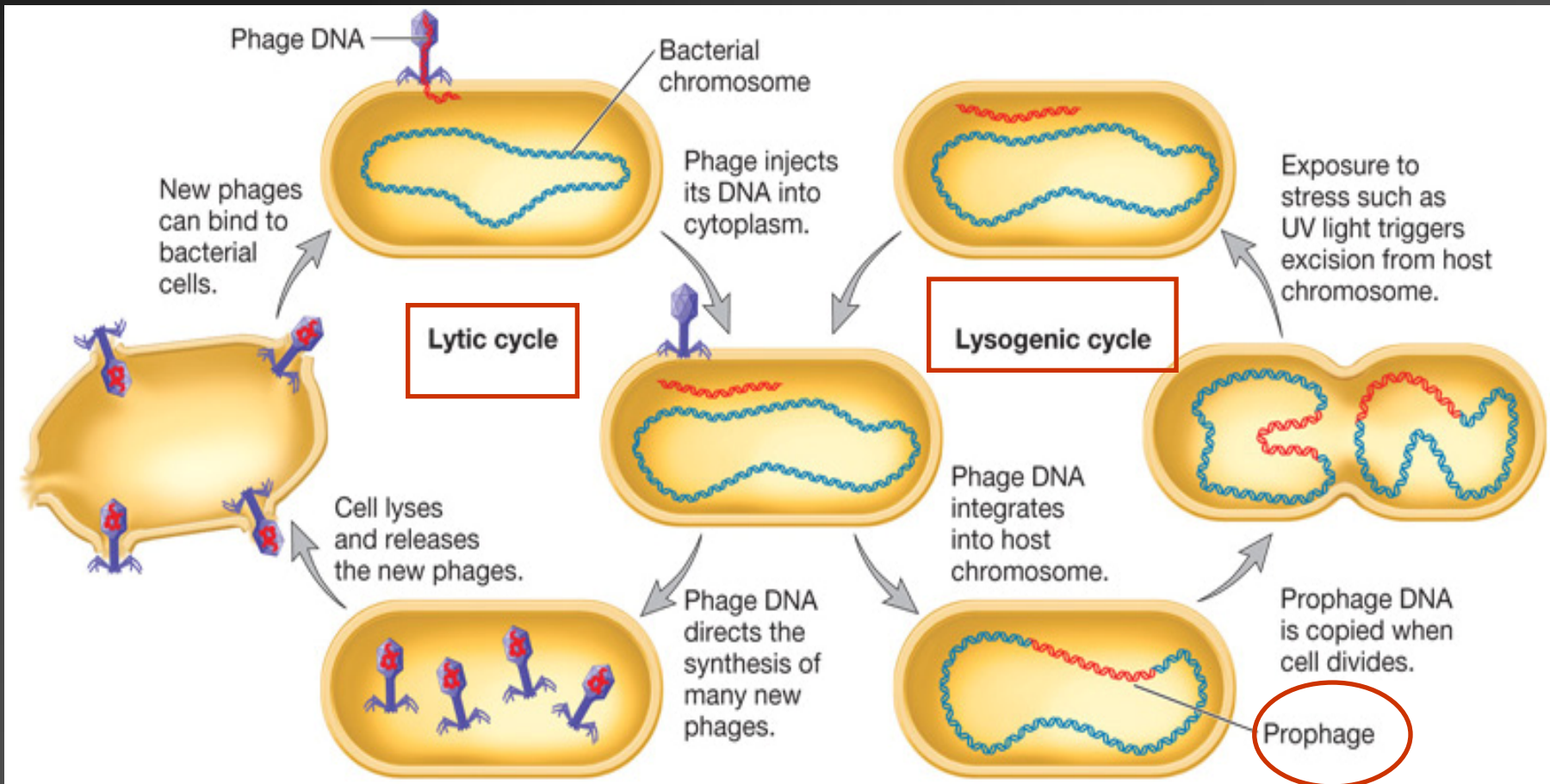


Fig 14.25

# Generalized transduction

- any part of bacterial genome can be transferred
- occurs during lytic cycle
- during viral assembly, fragments of host DNA mistakenly packaged into phage head

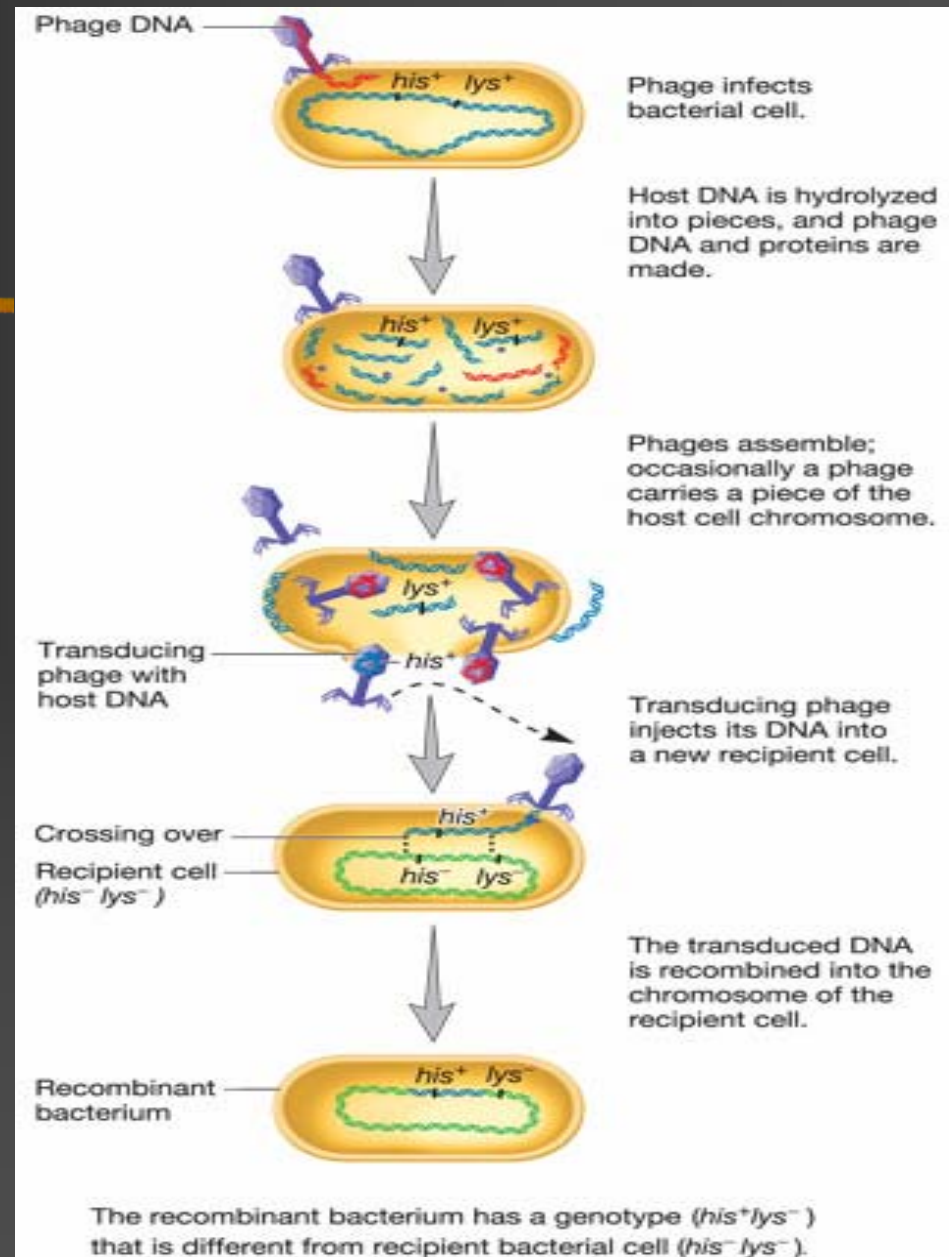


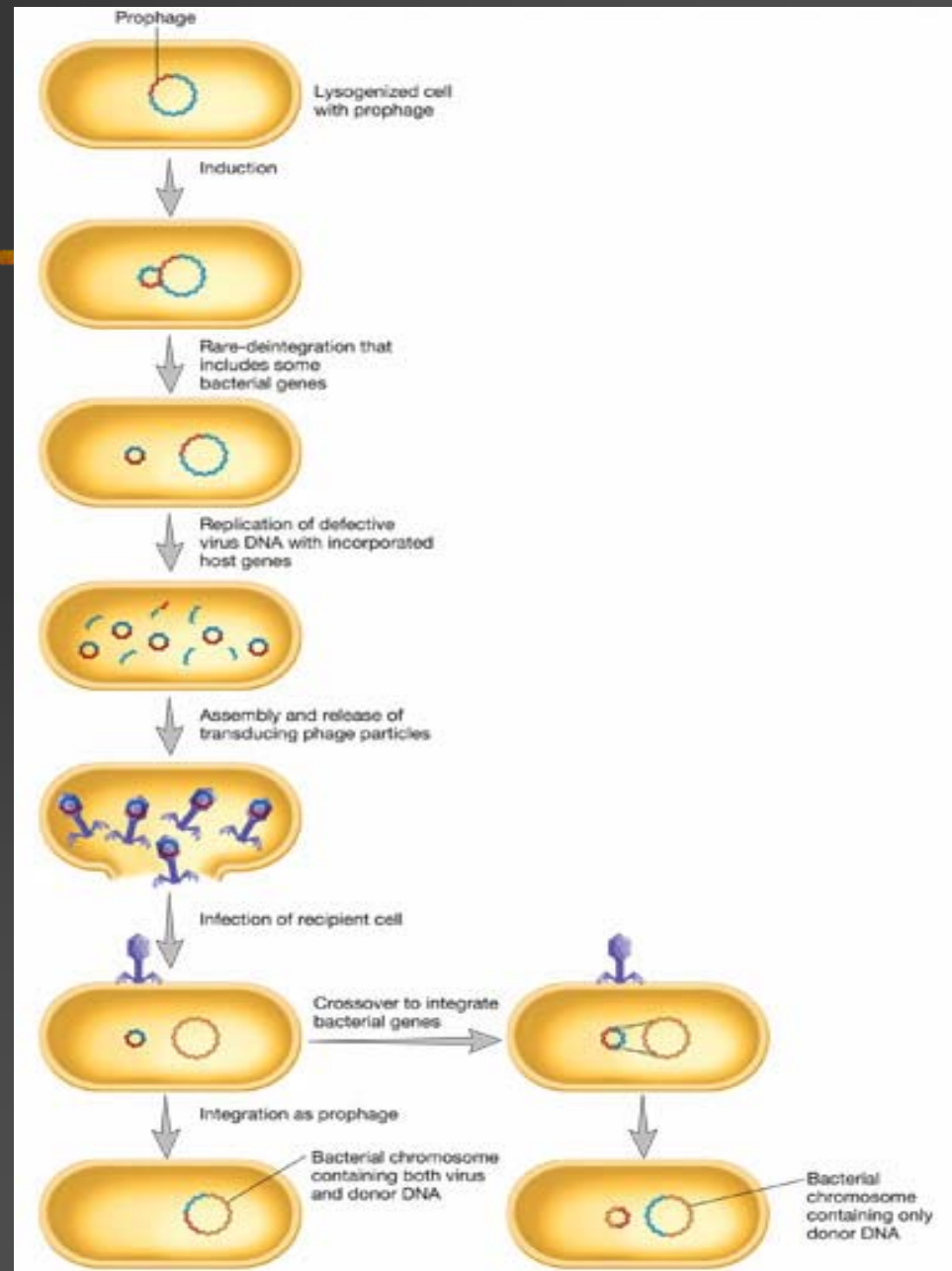
Figure 14.30



# Specialized Transduction

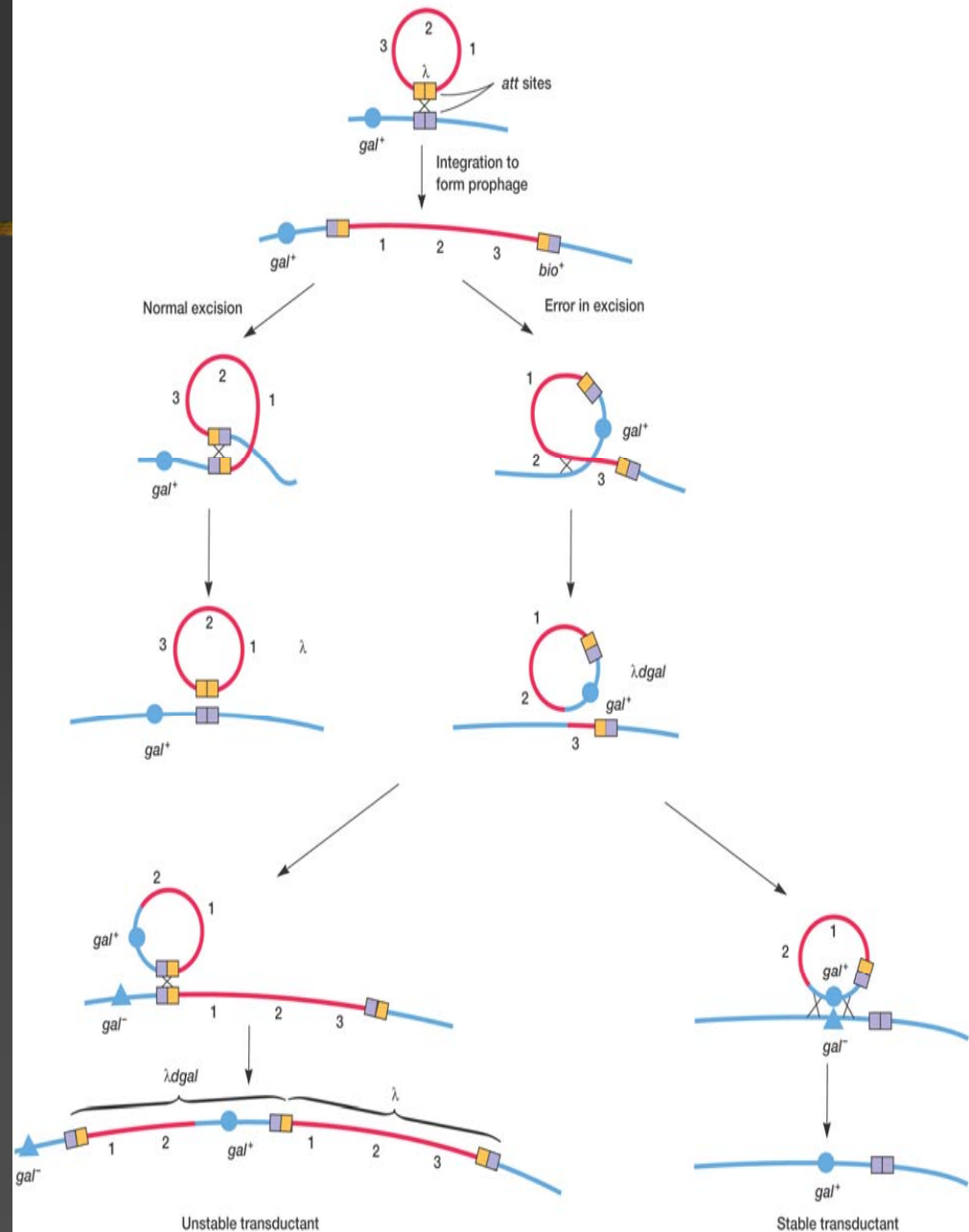
- carried out only by temperate phages that have established **lysogeny**
- only specific portion of bacterial genome is transferred
- occurs when **prophage** is incorrectly excised

Fig 14.31



# Transduction for phage Lamda

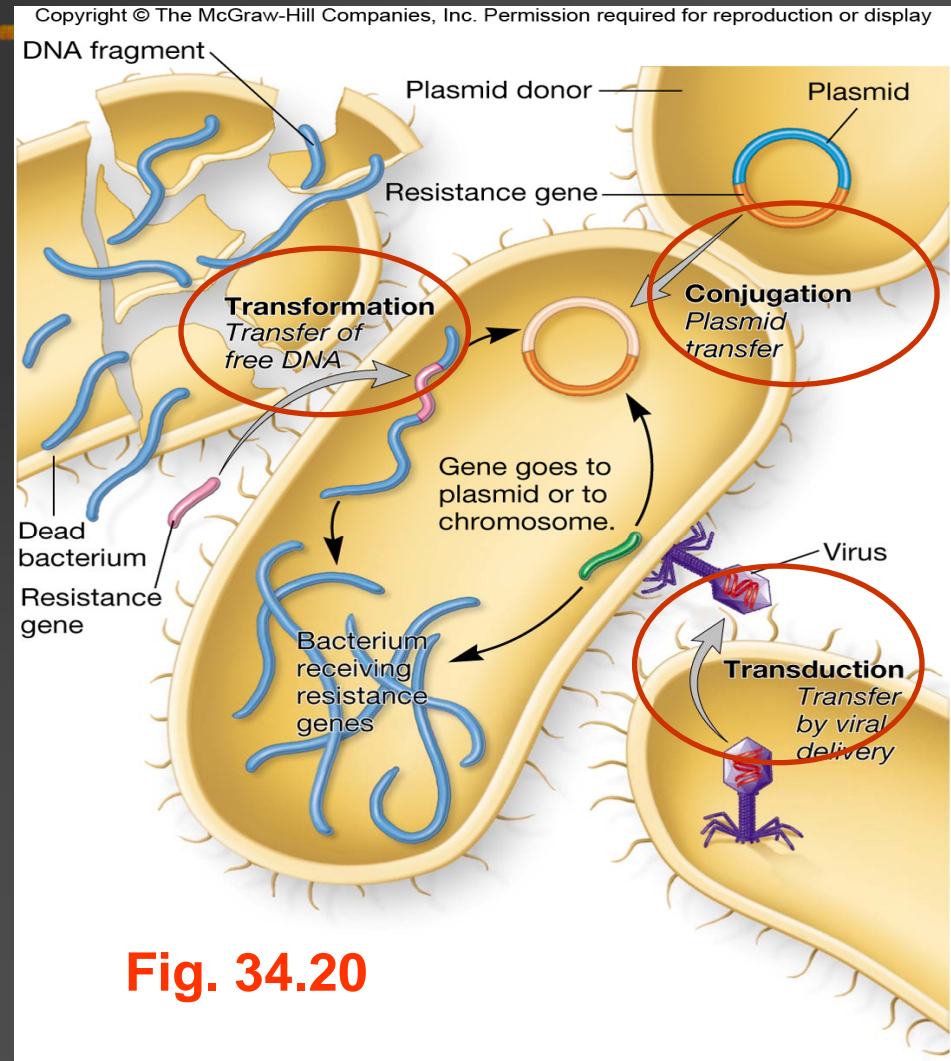
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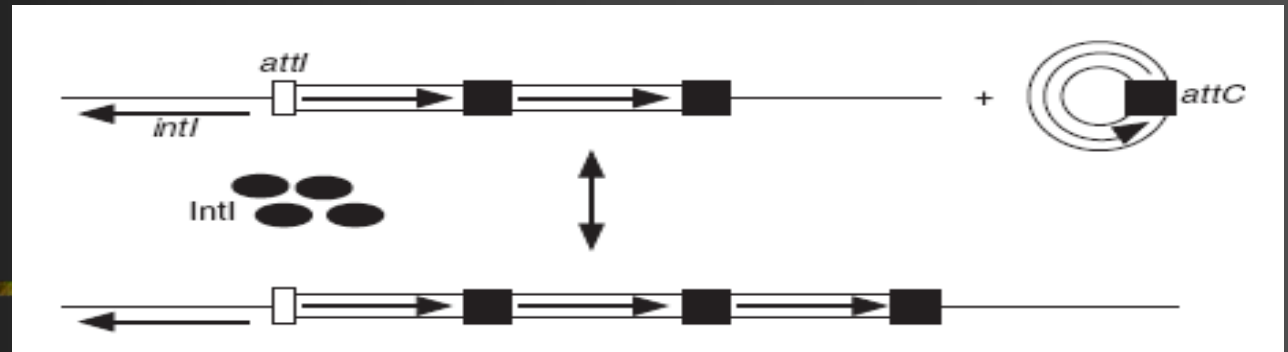


# Transmission of drug resistance

- The origin of drug resistance
  - R plasmid
    - Tn5, Tn21, Tn551
  - Integron
    - several resistance genes carried together as gene cassettes



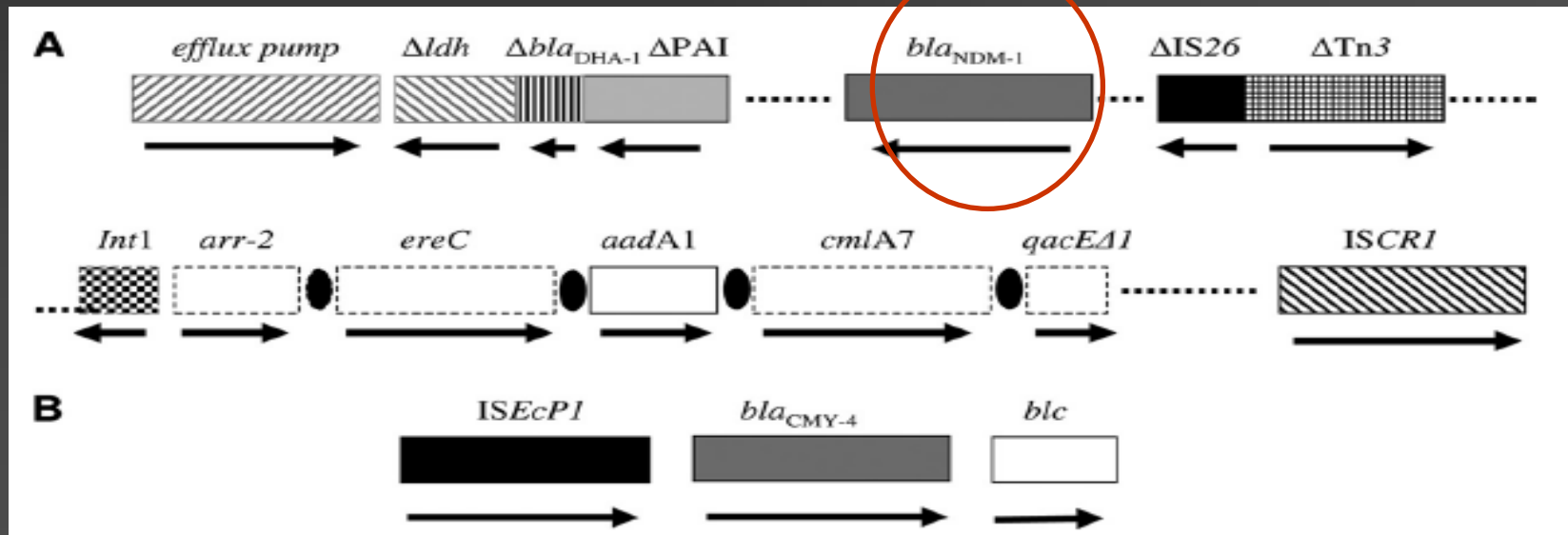
# Integron



- A gene cassette typically consists of little more than a single promoter-less gene and a recombination site
  - *intl* encodes an integrase of tryosine recombinase family
  - *attC* and *attI*
  - At least 130 different cassettes that carry known or predicted antibiotic resistance genes, along with many cassettes of unknown function

# Three characterized antibiotic resistance-conferring regions from *K. pneumoniae* 05-506

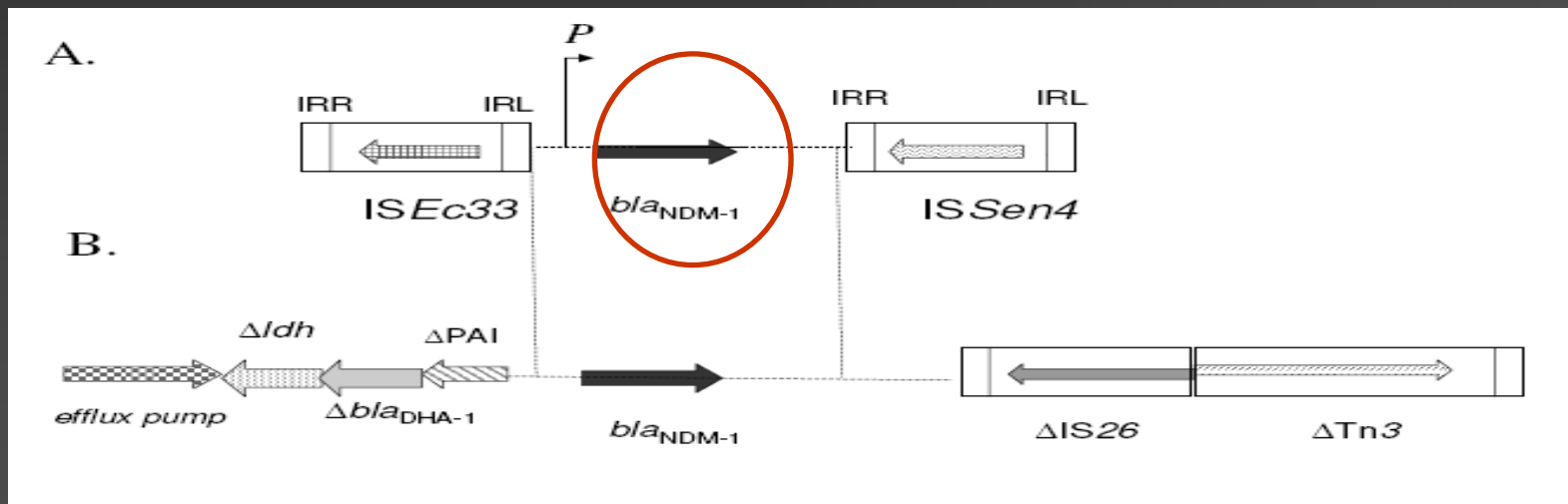
- A UTI (urinary tract infection) isolate of KPC strain 05-506, from a Swedish patient of Indian origin traveled to New Delhi, India, which carries a new subgroup of metallo-lactamase



Antimicrob. Agents Chemother., Dec. 2009, p. 5046–5054

# NDM-1 producing *E. coli*

- Emergence of NDM-1-producing multidrug resistant *Escherichia coli* in Australia





# Chapter 16

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## Microbial Genomics-1

# Genomics

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- study of molecular organization of genomes, their information content, and gene products they encode



# Determining DNA Sequences

- Sanger Method (1975)
  - uses dideoxynucleoside triphosphates (ddNTP) as chain terminator
- automated systems
  - use dideoxynucleotides labeled with fluorescent dyes

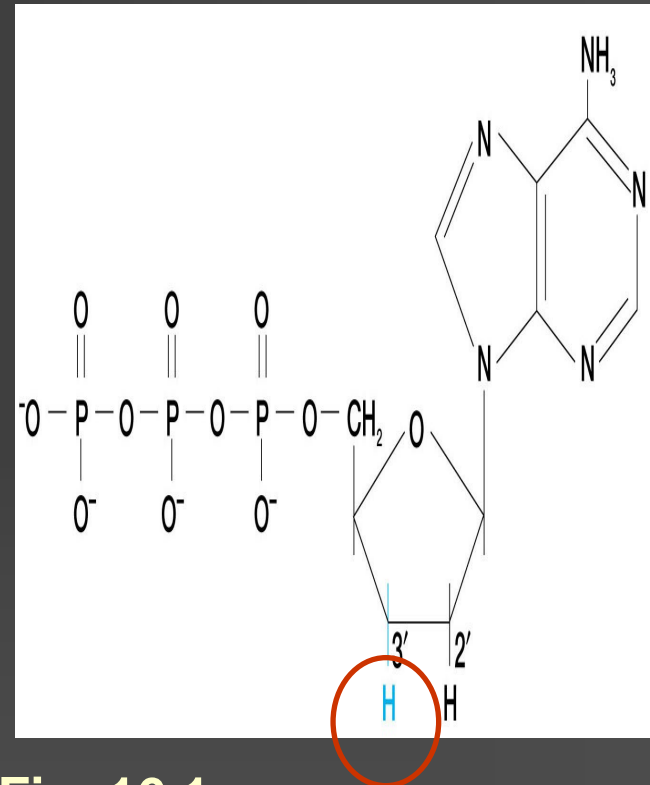
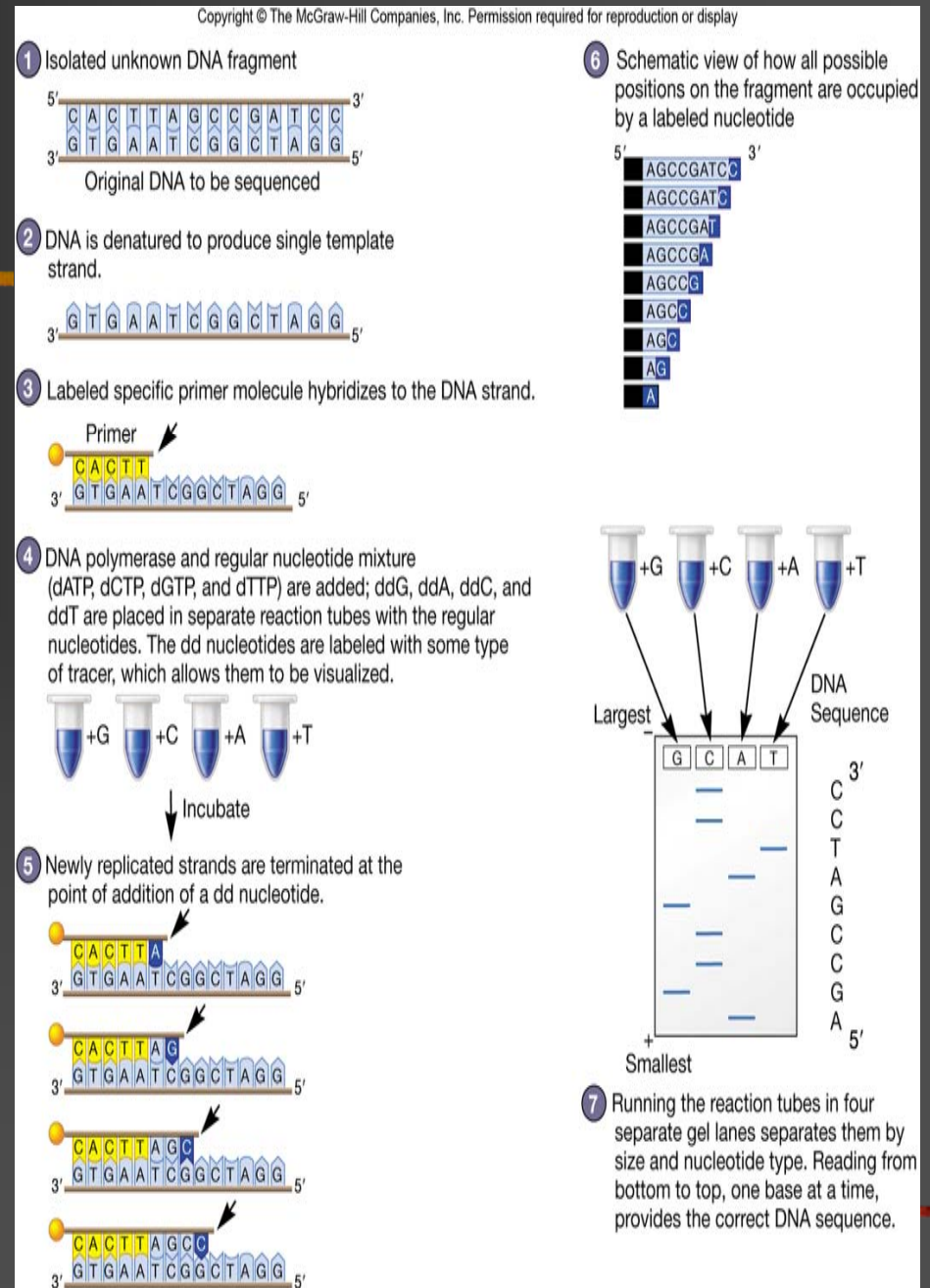


Fig. 16.1

# Chain termination method

- Four reactions
  - 1S DNA template
  - primer
  - DNA polymerase
  - 4 dNTP
  - one ddNTP
- random insertion of ddNTP generates different lengths of DNA fragments
- fragments separated electrophoretically
- sequence read

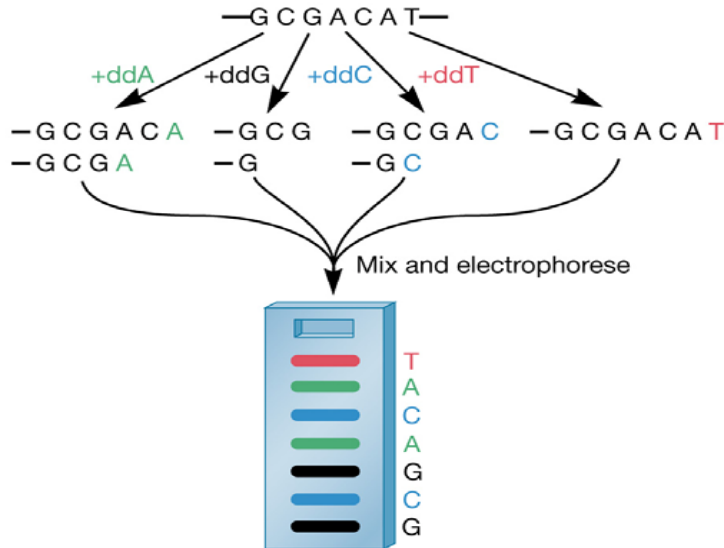
Fig. 16.2



# Automated Sanger DNA sequencing

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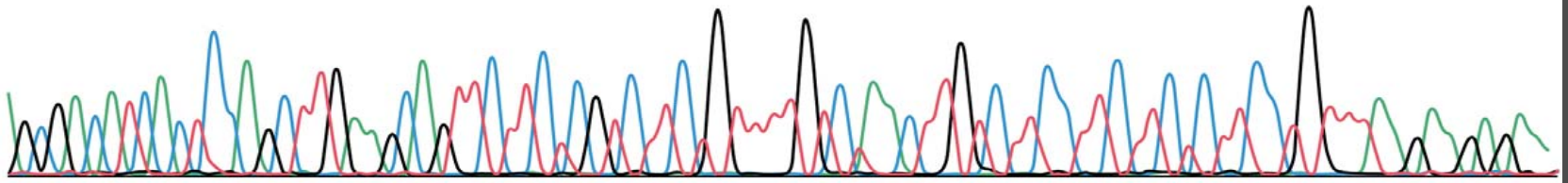
Fig. 16.3



(a)

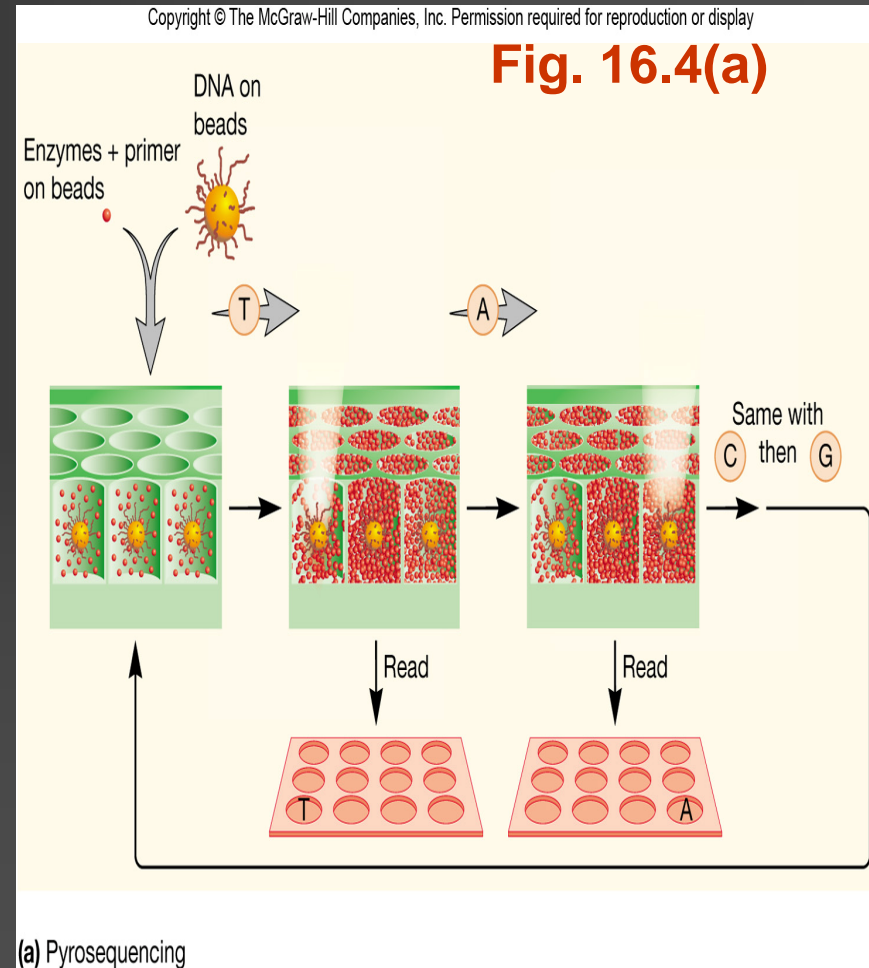
GCGACATCACTCCAGCTTGAAGCAGTTCTTCTCGTCTTCTGTTTTGTCTAACTTGTCTTCTCTTCTCTTCTCTTCTGTTTAAGAAGAGAA  
500 510 520 530 540 550 560 570 580

(b)



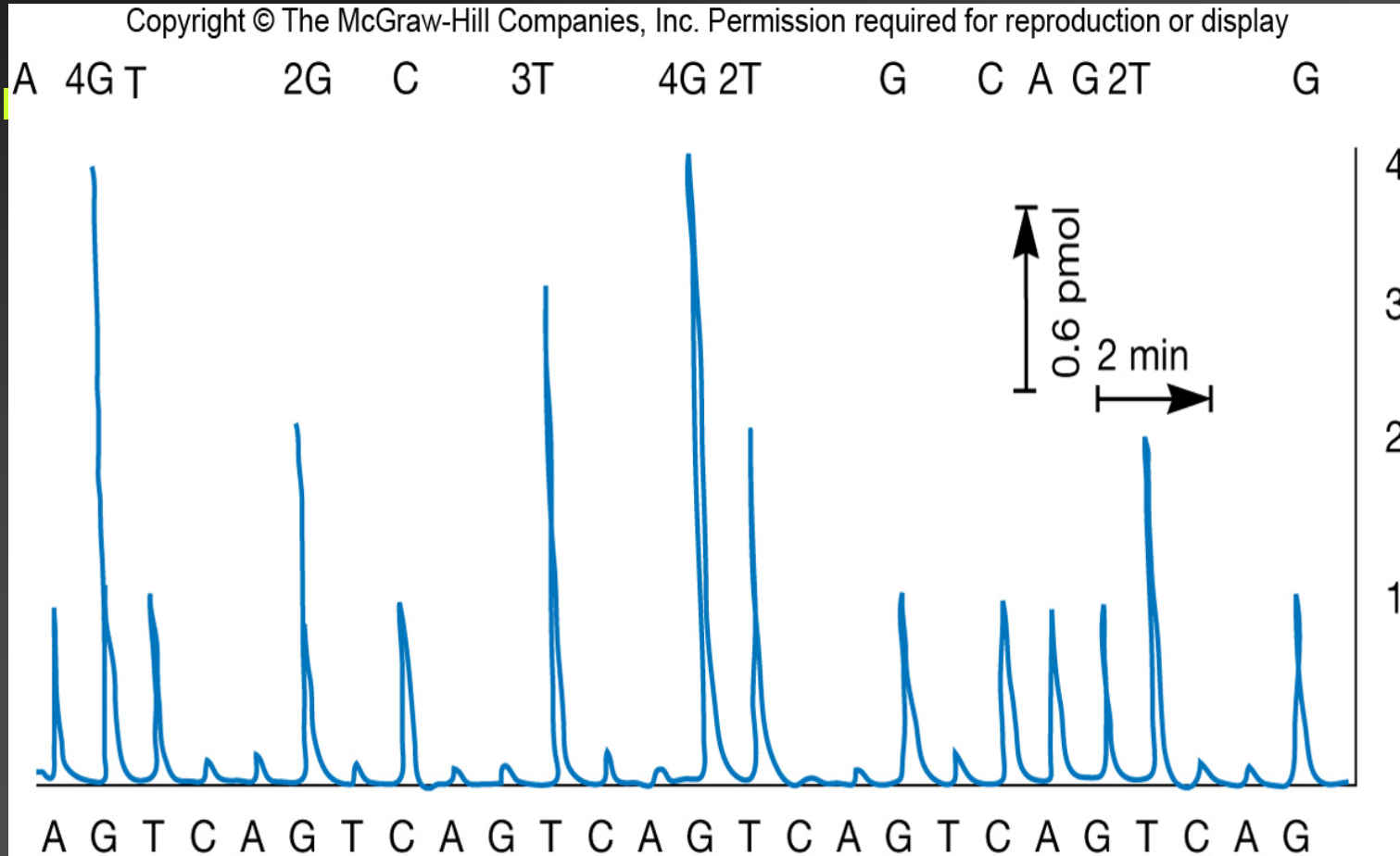
# Post-Sanger DNA sequencing- pyrosequencing

- Each bead coated with PCR-amplified chromosomal fragments (300-500 bp)
- pyrophosphatase sequencing
  - $(DNA)_n + dNTP \rightarrow (DNA)_{n+1} + PPI$  by DNA polymerase
  - $PPI + APS$  (adenosine phosphosulfate)  $\rightarrow ATP + SO_4^{2-}$  by ATP sulfurylase
  - $ATP + luciferin + O_2 \rightarrow AMP + ppi + oxyluciferin + CO_2 + \text{light}$  by luciferase



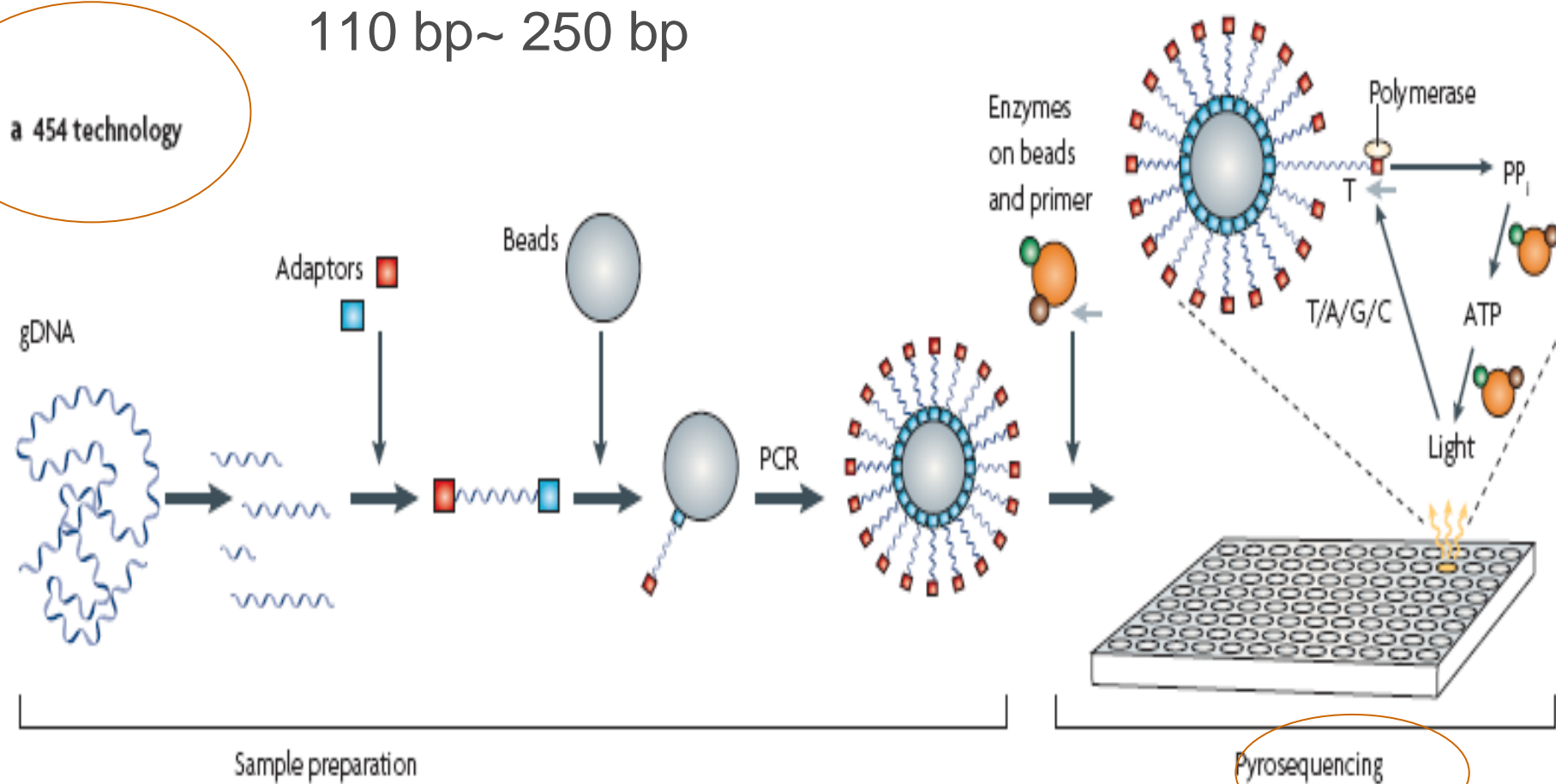
# A Pyrogram

Fig. 16.5



# Roche 454 Genome Sequencing

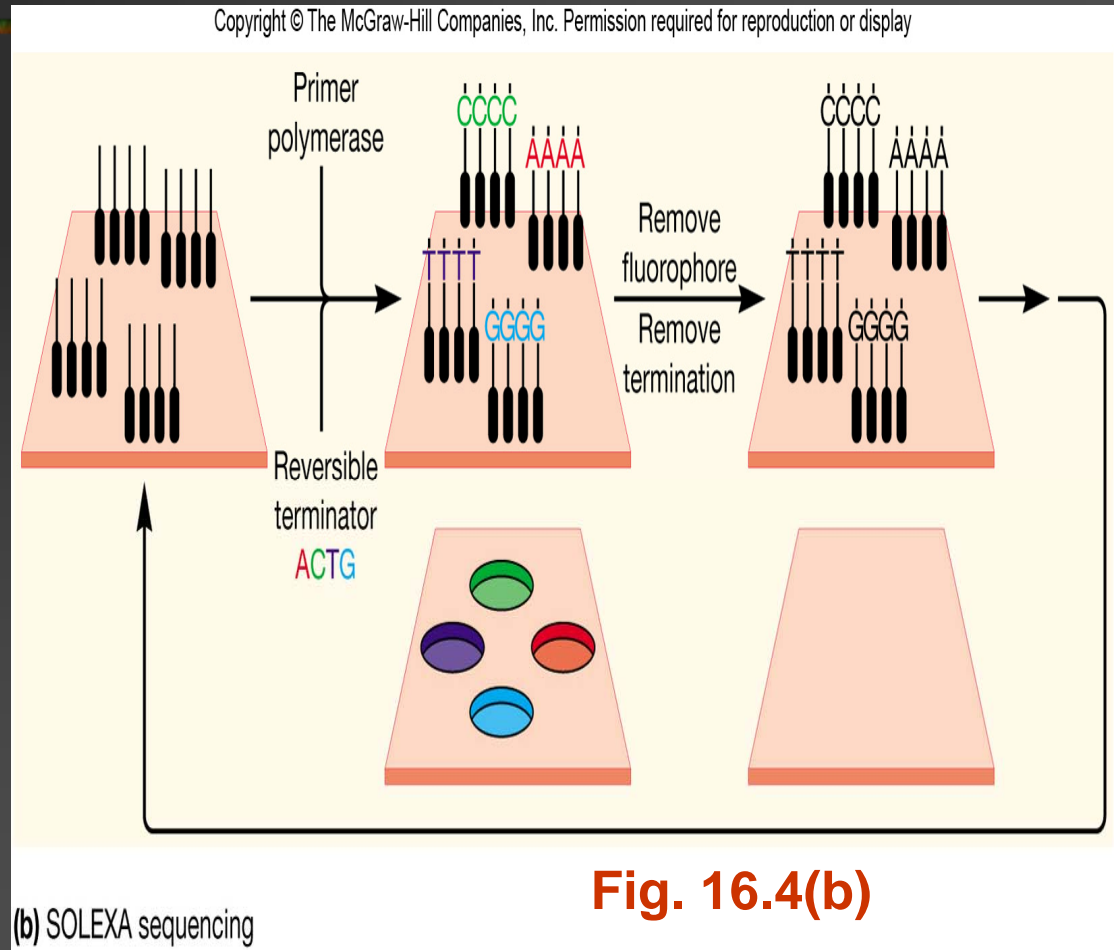
Nature Rev  
Microbiology  
(2008) May





# SOLEXA sequencing

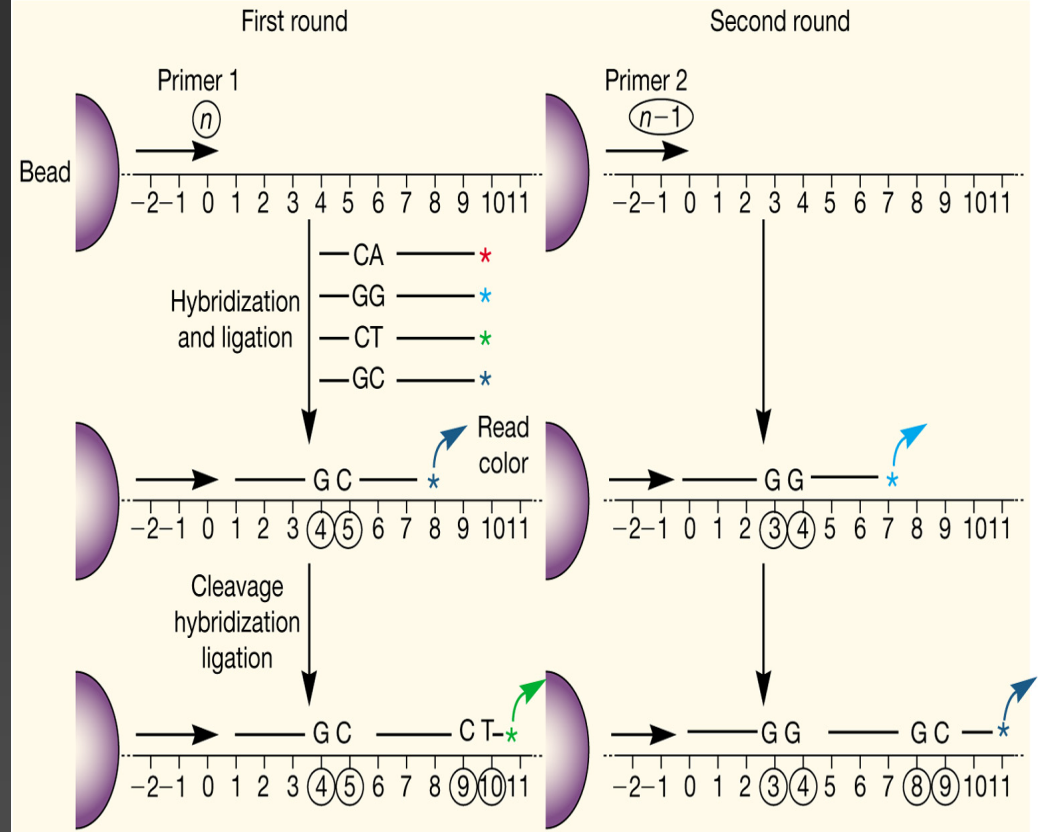
- A glass slide attached with the PCR amplification of immobilized DNA fragments
- Sanger-type sequencing
  - 4 fluorophore terminators



# SOLiD sequencing

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- Supported oligo ligation detection
- The DNA fragments are lengthened by
  - the addition of a short piece of DNA (the anchor primer)
  - eight-base oligonucleotides (by every possible combination of A-T-C-G) and each has a fluorescently labeled A, T, C, or G in the 4th and 5th positions
  - Ligase
  - Exciting with laser light



(c) Sequencing by ligation (SOLiD technology)

**Fig. 16.4(c)**

# Whole-genome shotgun sequencing

- Library construction to provide the templates
  - genomic clones
- random sequencing
- fragments alignment and gap closure
  - **Contig**- a set of larger, contiguous nucleotide sequences

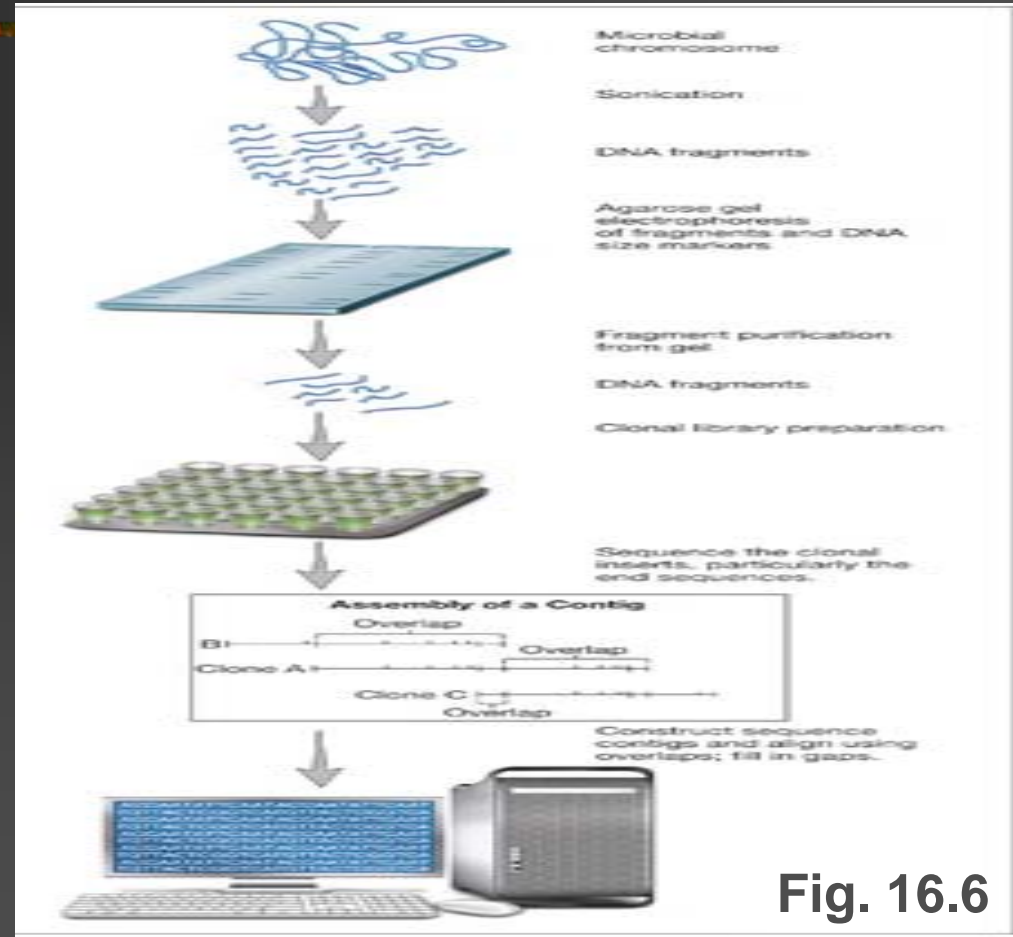


Fig. 16.6

- Editing (proofread)
  - 1000 kb/ 24 h

1995, J. Craig Venter and Hamilton Smith  
(TIGR: the institute of genomic research)

# Single-Cell genomic sequencing

- Multiple strand displacement (NDA)
  - DNA from single cell (down to  $10^{-15}$  g)
  - random primers (hexamers)
  - phage phi29 DNA polymerase
  - dNTPs
- Many new strands (12 kb~100 kb) are synthesized → cloning → sequencing

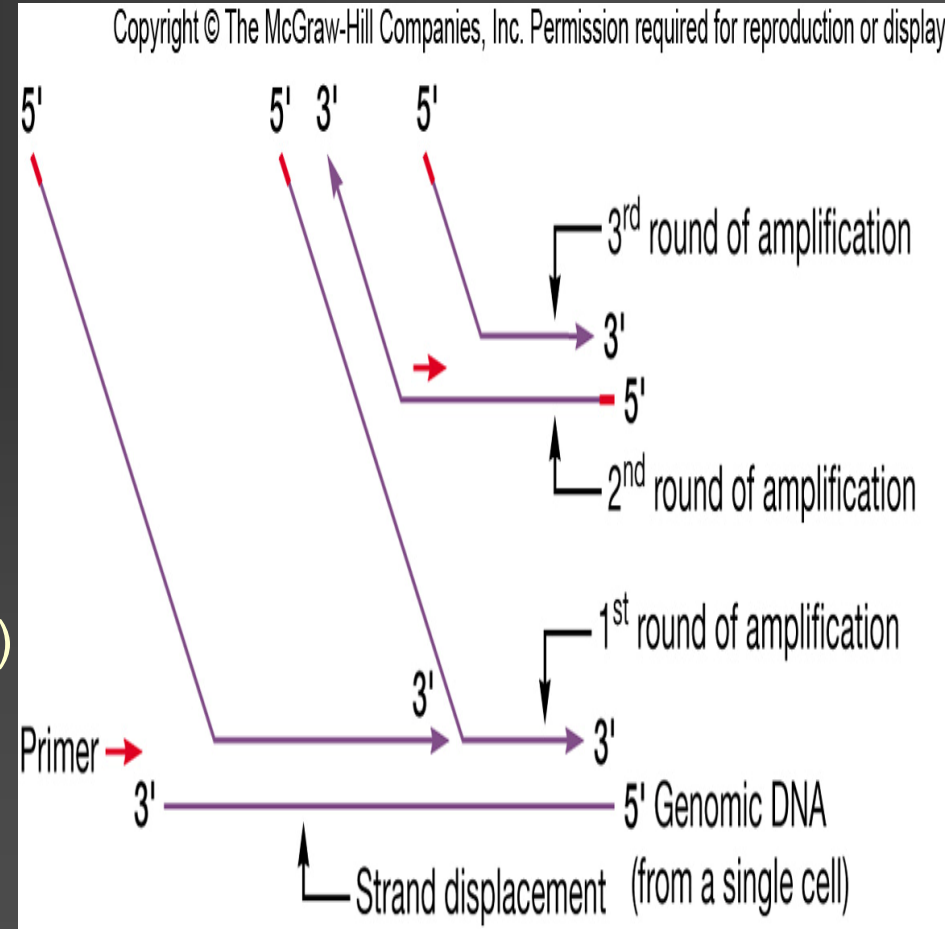


Fig. 16.7

# Bioinformatics

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- Combine Biology, mathematics, computer science, and Statistics to generate data on genome content, structure, and arrangement
  - also provides data on protein structure and function
  - DNA sequence data stored in large **databases**
    - International Nucleic Acid Sequence Data Library (**GenBank**)
  - analysis of genome data using computers
    - *in silico* analysis

# Genome annotation

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- To determine location of genes and nature of genes or presumed genes
  - identifies **open reading frame** (ORF) in genome
    - a reading frame  $> 100$  codons that is not interrupted by a stop codon
    - there is an apparent ribosomal binding site at the 5' end and terminator sequences at the 3' end
  - To assign tentative function of gene
-



# Finding potential protein coding genes

## - CDS (coding sequences)

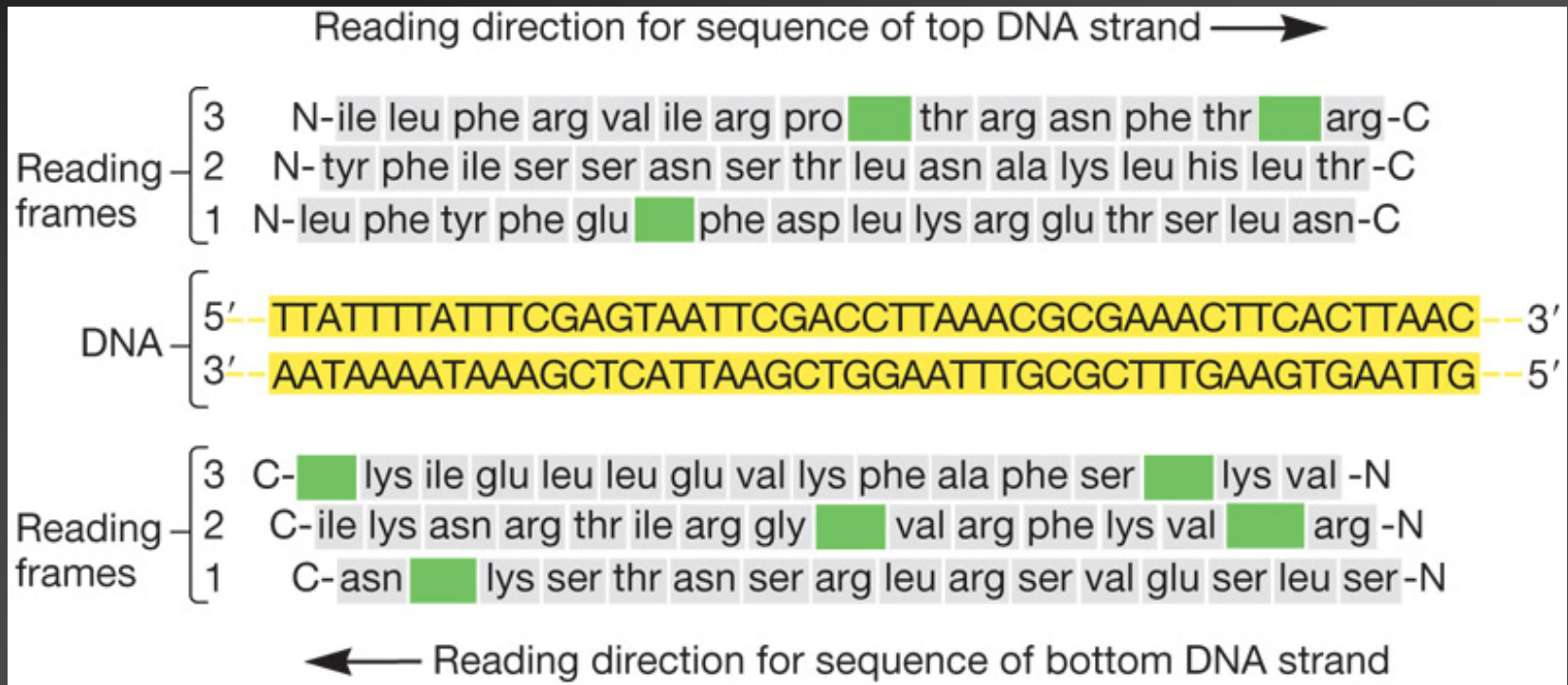


Fig 16.8

# Alignment of the conserved regions



Fig 16.9 Pfam database



# Chapter 16

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## 16. Microbial Genomics- 2

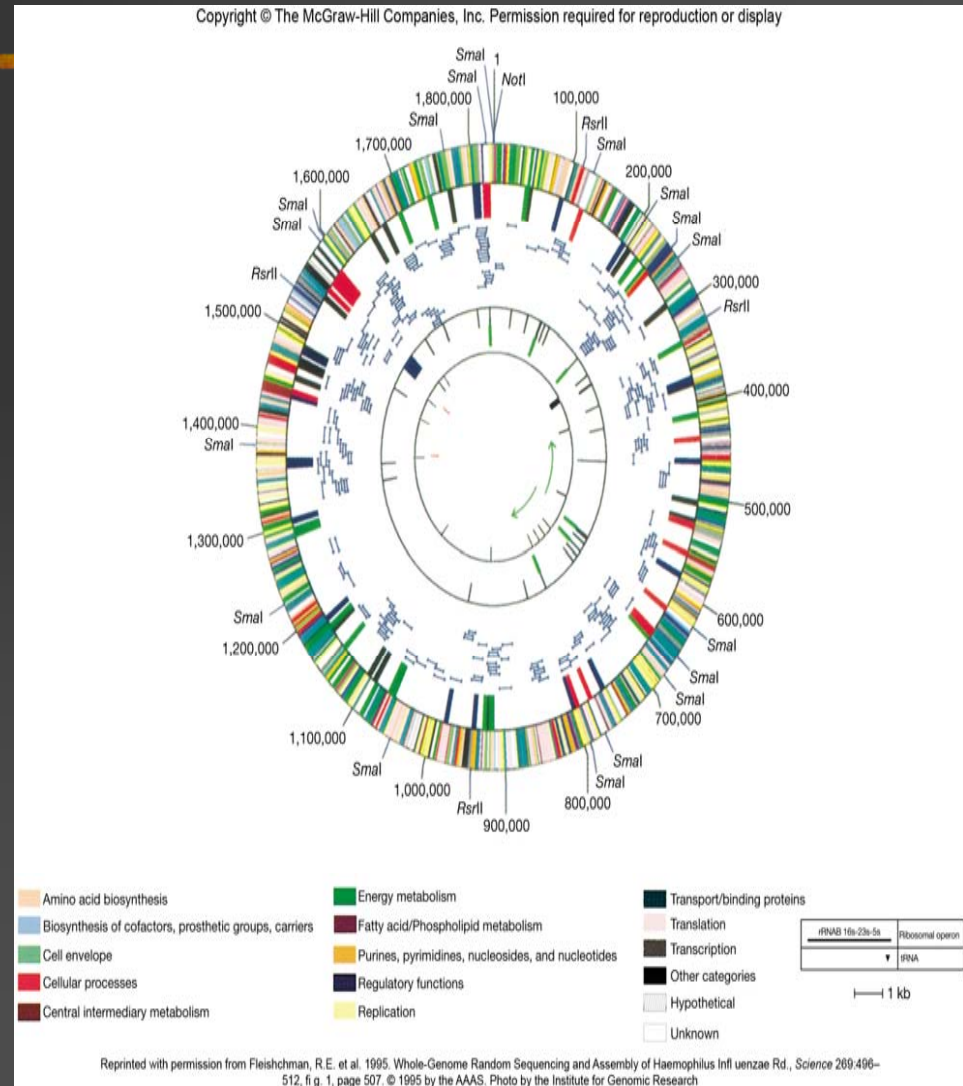
11-17-2010

# Functional genomics

- Determination of how genome works
  - from alignment of gene sequences
    - **paralogs** – genes arose from gene duplication
    - **orthologs** – genes very similar and are predicted to have same function
  - involves analysis of translated amino acid sequence of presumed genes to understand protein structure and function
    - **motif**, a short pattern of amino acids, may represent a functional unit within the protein, such as the active site

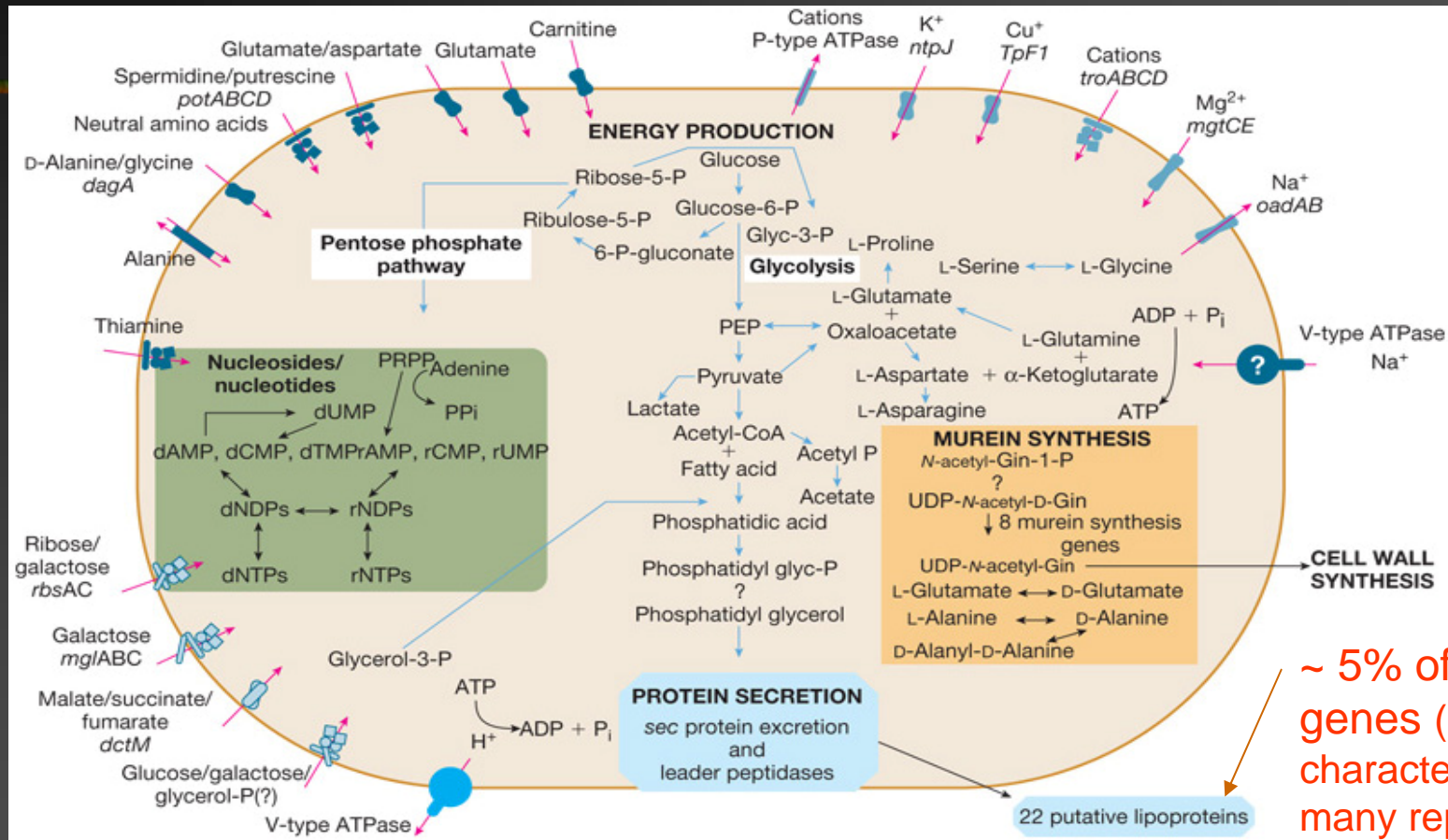
# Physical map of the *Haemophilus influenzae* genome

- 1st annotated genome
  - Color-code genes
    - 1/3 of unknown function (white regions)
    - 65 regulatory genes (dark blue)
- > 700 complete bacterial and archaeal sequences now available





# Treponema pallidum- syphilis 梅毒



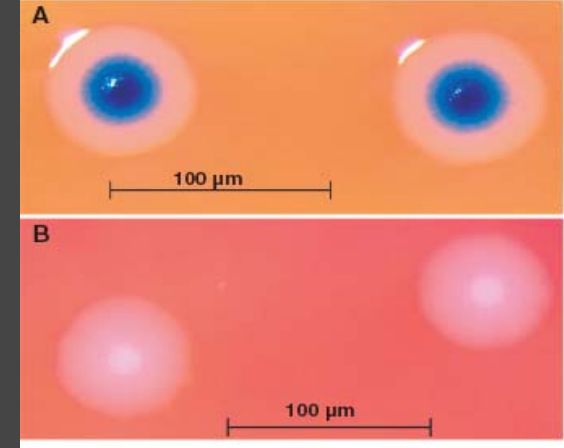
~ 5% of the genes (some characterized by many repetitive sequences → recombination → generate new surface proteins)

Figure 16.11

metabolically crippled- No TCA cycle and respiratory electron transport



# First synthetic genome



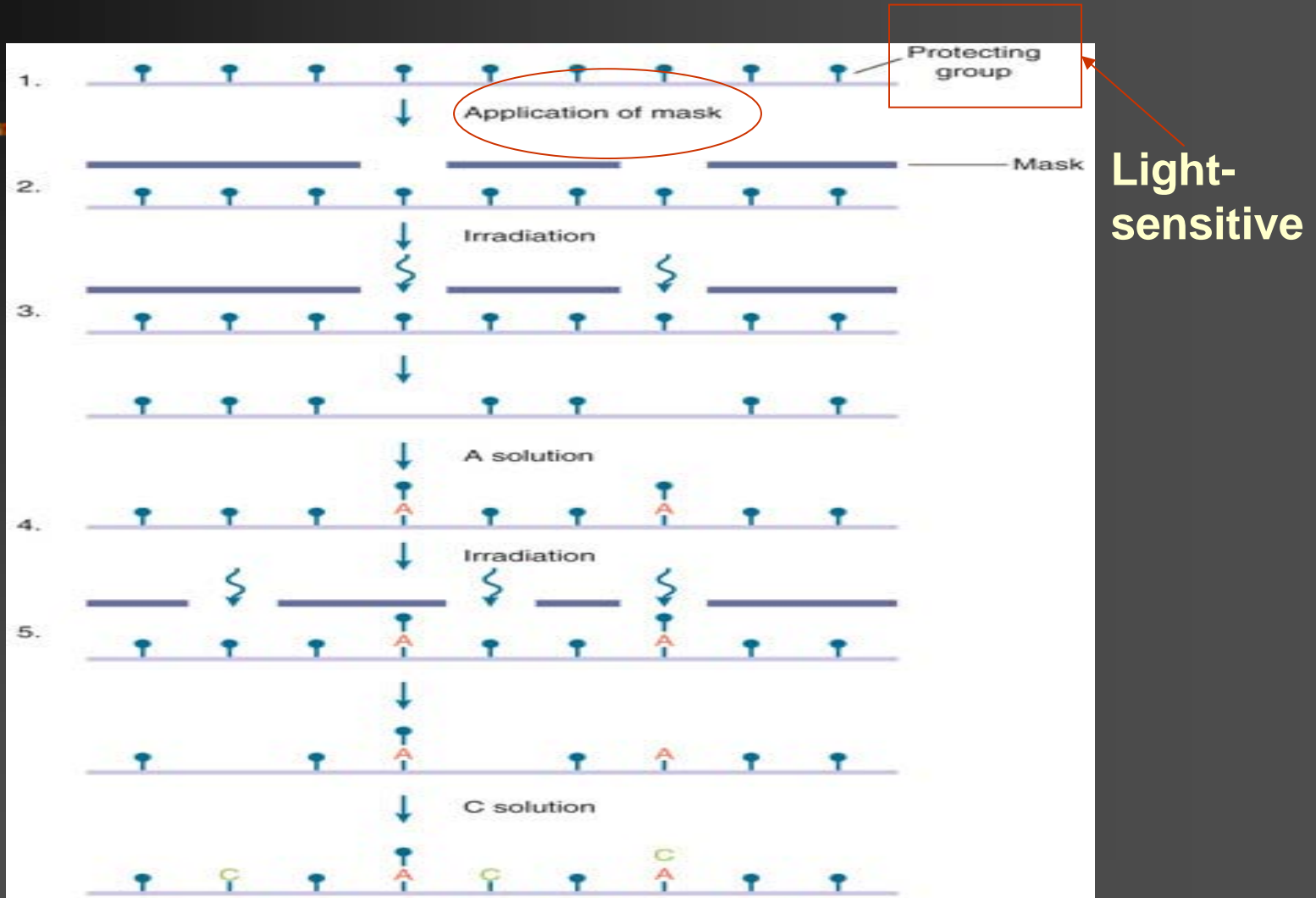
- *Mycoplasma mycoides* JCVI-syn1.0
  - 1.08 million bp synthesized chromosome of a modified *M. mycoides* genome at the J. C. Venter Institute (San Diego, CA, and Rockville, MD, USA)
  - ~ 1 kb → 10 kb in *E. coli* → 100 kb in yeast → assembly of 10 X ~100 kb fragments in yeast → mycoplasma
  - The new cells have expected phenotypic properties and are capable of continuous self-replication (>30 divisions) and making new set of proteins.

# DNA microarray analysis (Gene Chips)

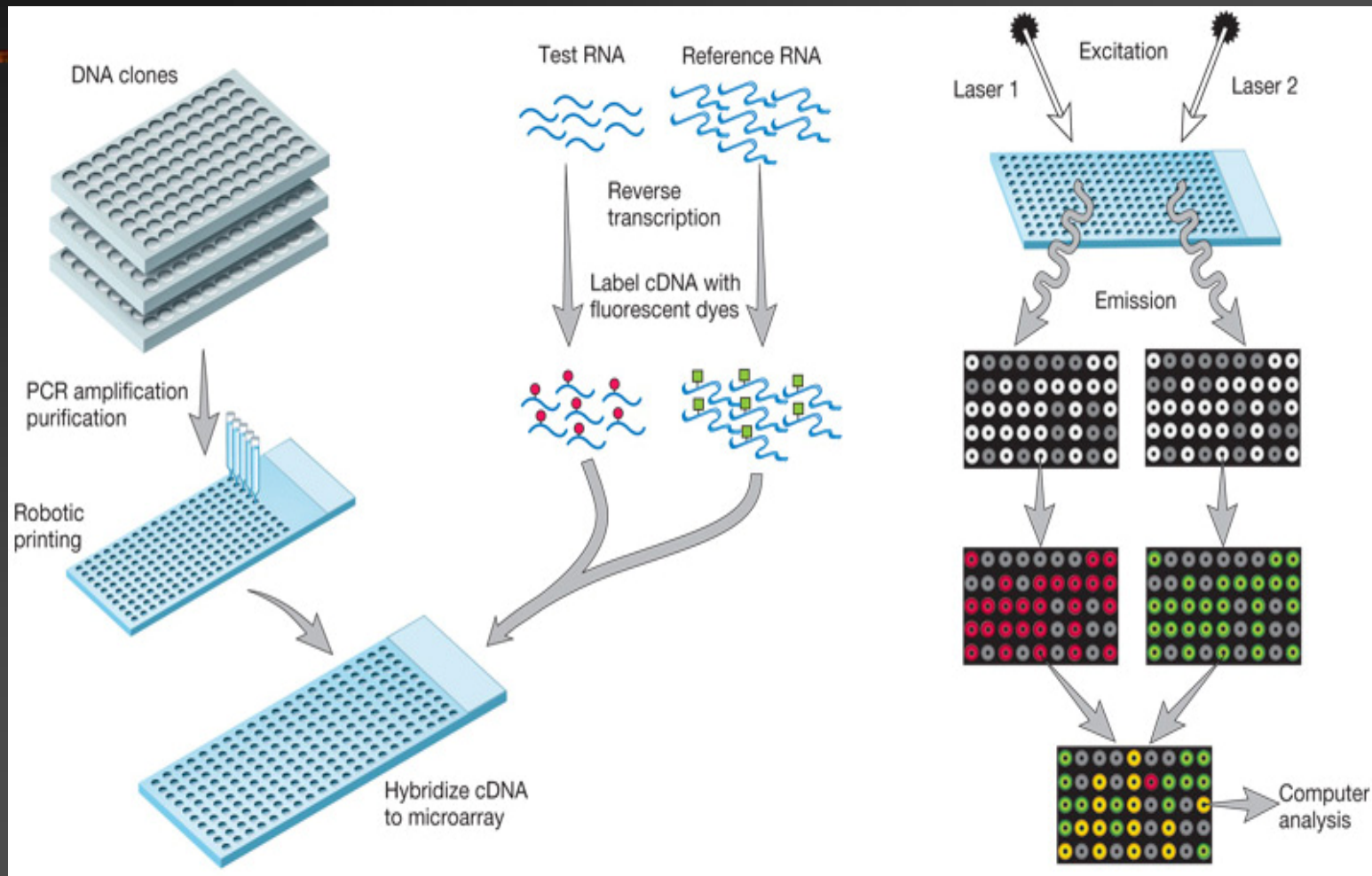
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- Determine gene expression at a specific time
  - **spotted arrays**
    - prepared by robotic application of DNA probe
    - each DNA **probe** represents a single gene or ORF
      - PCR product, cDNA or oligonucleotide
      - oligonucleotide probes from eukaryotes-
        - **expressed sequence tags (EST)**
  - **photolithography**

# Construction of DNA chip- photolithography



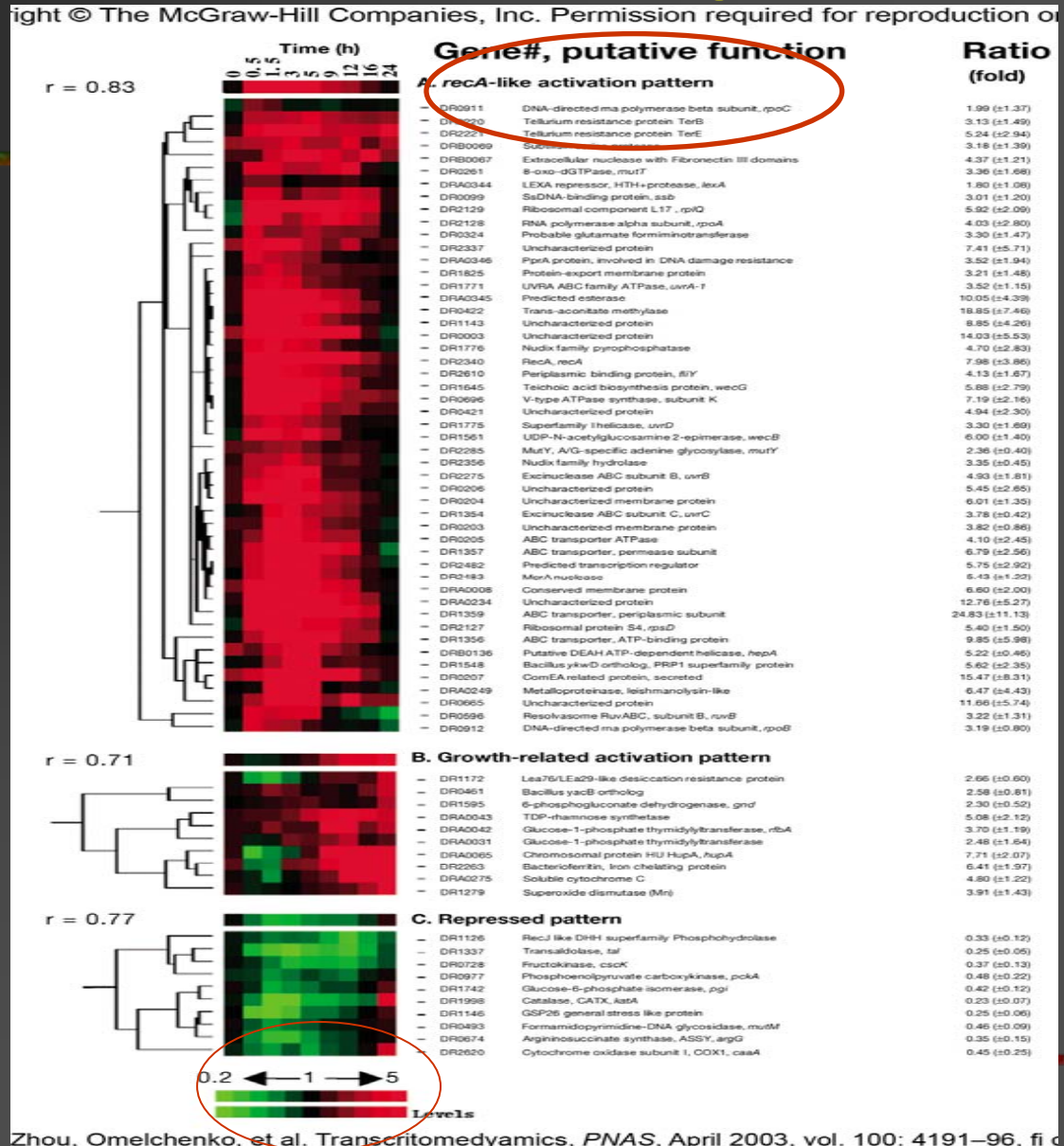
# Monitoring changes of gene expression



# Hierarchical cluster analysis of gene expression

Analysis of gene expression of *D. radiodurans* following exposure to  $\gamma$ -radiation

- Each group of genes has been scored for relatedness
- A tree has been generated with indication of **correlation coefficient** ( $r$  value)
- Genes **induced** (red) or **repressed** (green) after radiation



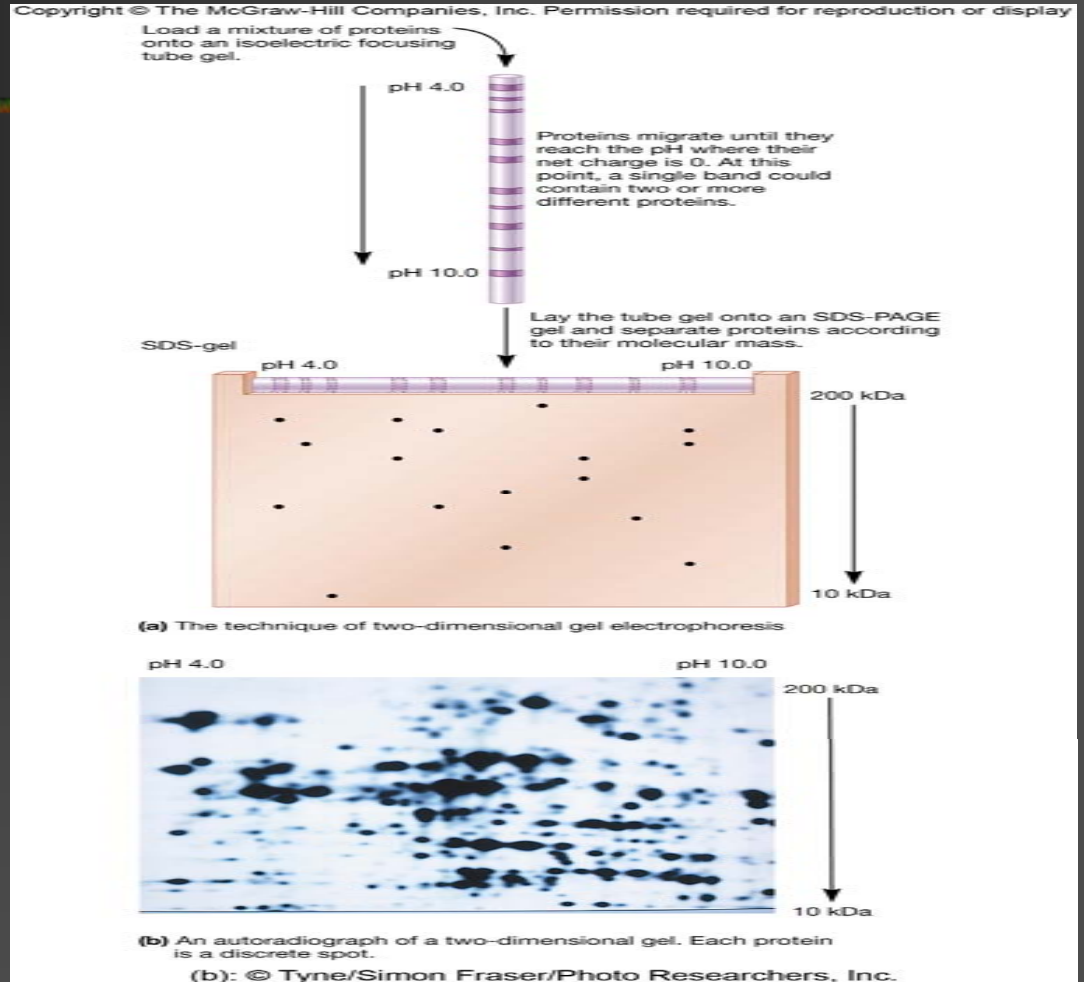
# Proteomics

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- The study of the **proteome** (the entire collection of proteins )
  - often analyzed by two-dimensional gel electrophoresis
  - **Functional proteomics** to determine what is actually happening in cell proteome
  - **Structural proteomics** to resolve 3 dimensional structure of the proteins
    - **protein modeling** ( on the basis of the assumption that proteins folds into a limited number of shapes)



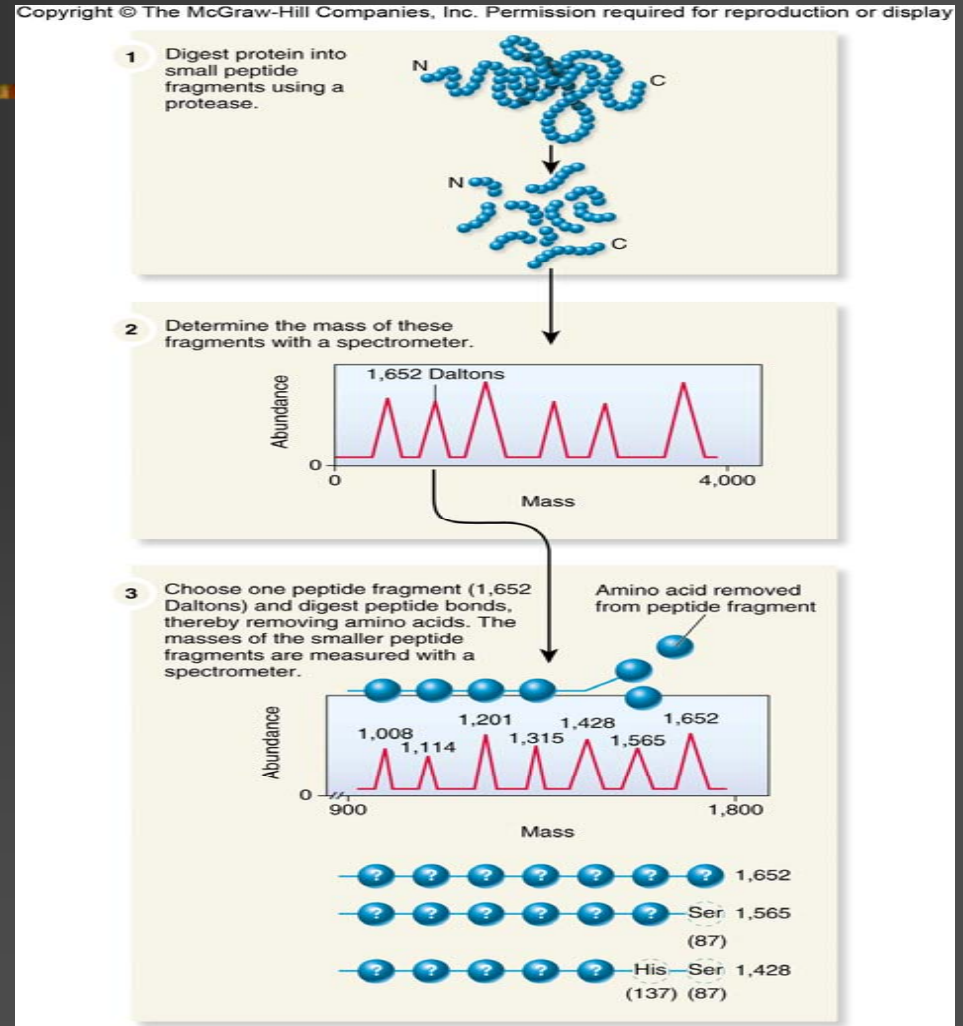
# Two-dimensional gel electrophoresis





# Tandem Mass Spectrometry

- unknown spot from 2-D gel is cut and cleaved
- fragments are analyzed by mass spectrometer
- mass of fragments is plotted
- protein tentatively identified from probable amino acid composition

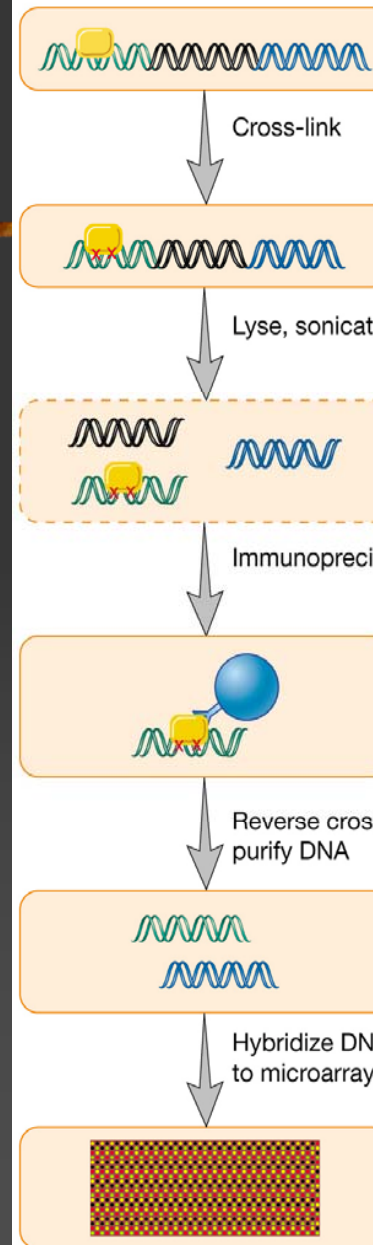


**Fig 16.15**

# ChIP-Chip analysis

- probing DNA-protein interaction
  - **EMSA** (electrophoretic mobility shift assay) or called gel mobility shift assay
  - **ChIP**- chromatin immunoprecipitation
    - search for protein binding DNA

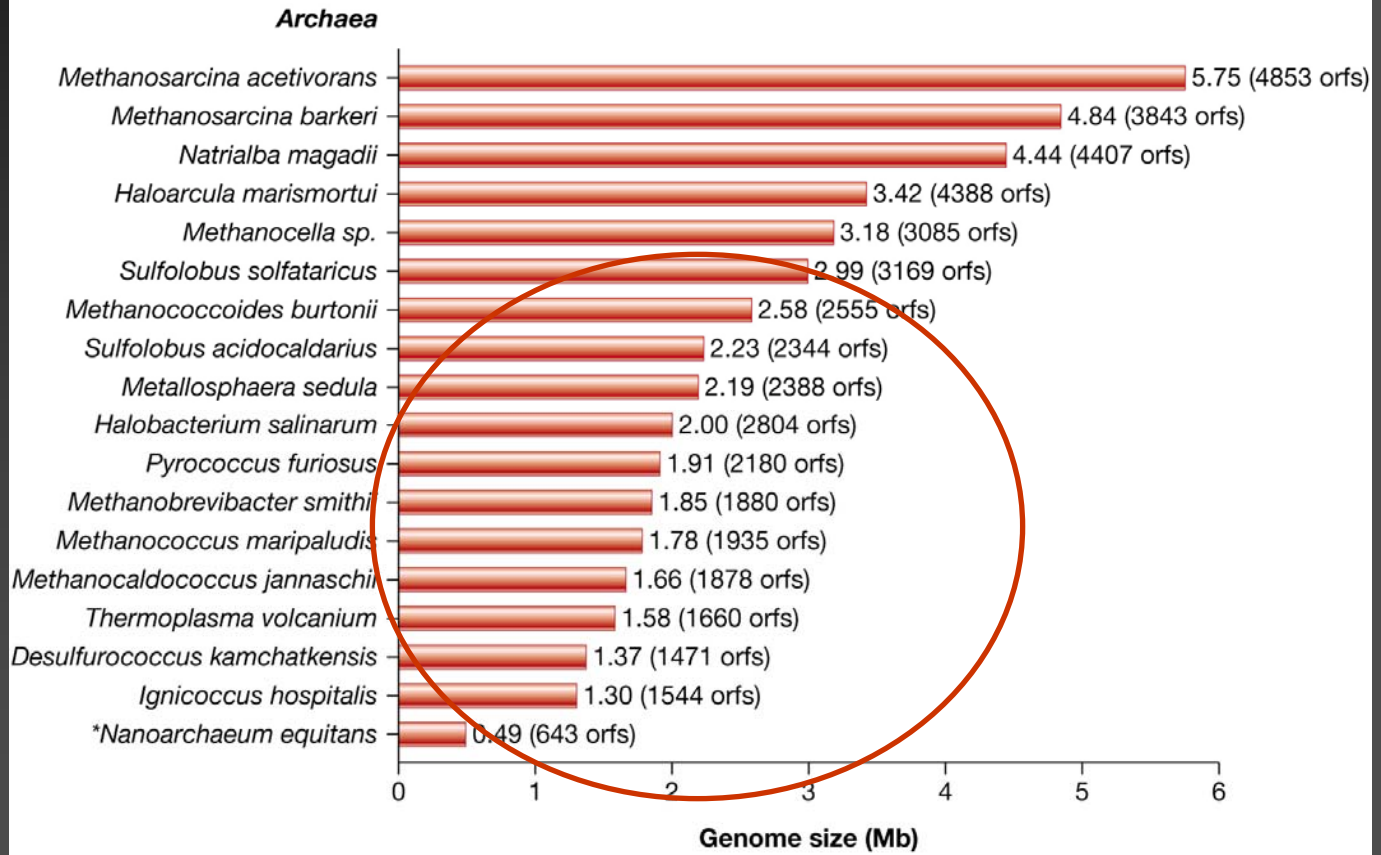
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- 1 Microbial cells are grown under the conditions of interest.
- 2 A cross-linking agent such as formaldehyde is added so that the protein is stably attached to the DNA.
- 3 The cells are broken and the DNA is fragmented by sound waves, a process known as sonication.
- 4 Antibodies are added and bind to the target protein. This increases the weight of the protein-DNA complex, so it can be selectively precipitated, a process known as immunoprecipitation.
- 5 Cross-linking is reversed by heating, which also denatures the DNA.
- 6 The DNA is fluorescently labeled and hybridized to a microarray for identification.

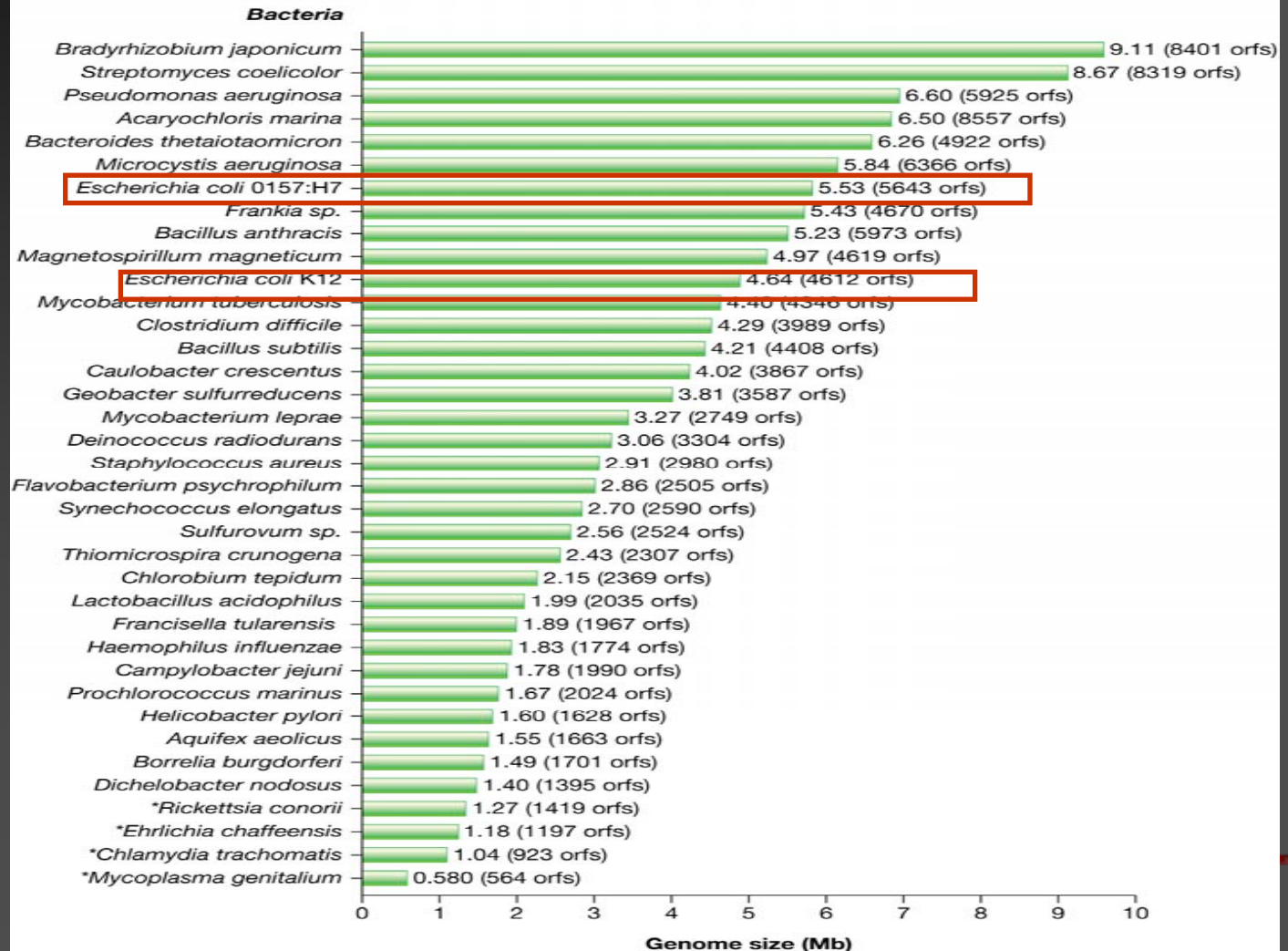
# Comparative Genomics- Archaea genome size

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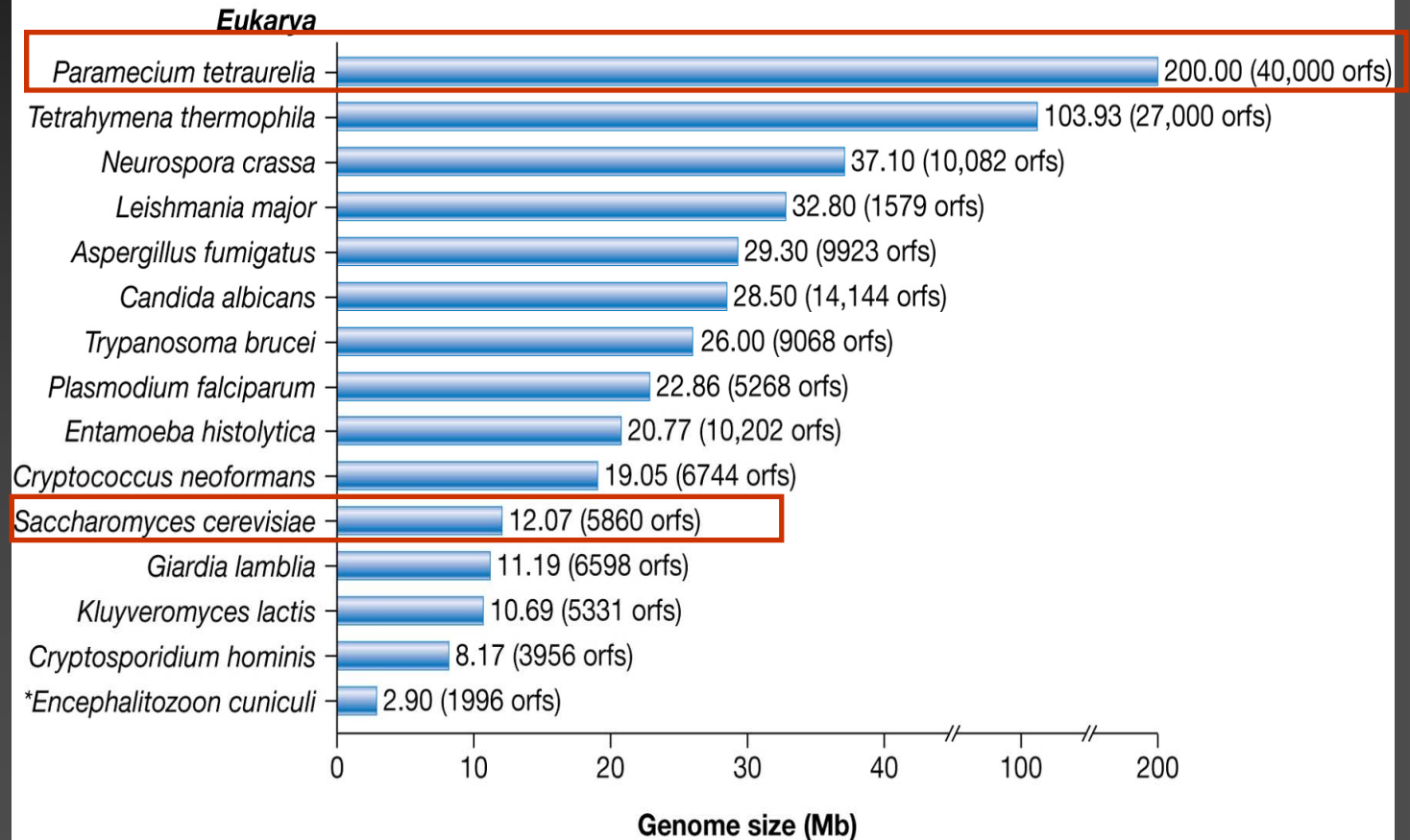
# Comparative Genomics- Bacteria genome size

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# Comparative Genomics- *Eukarya* genome size

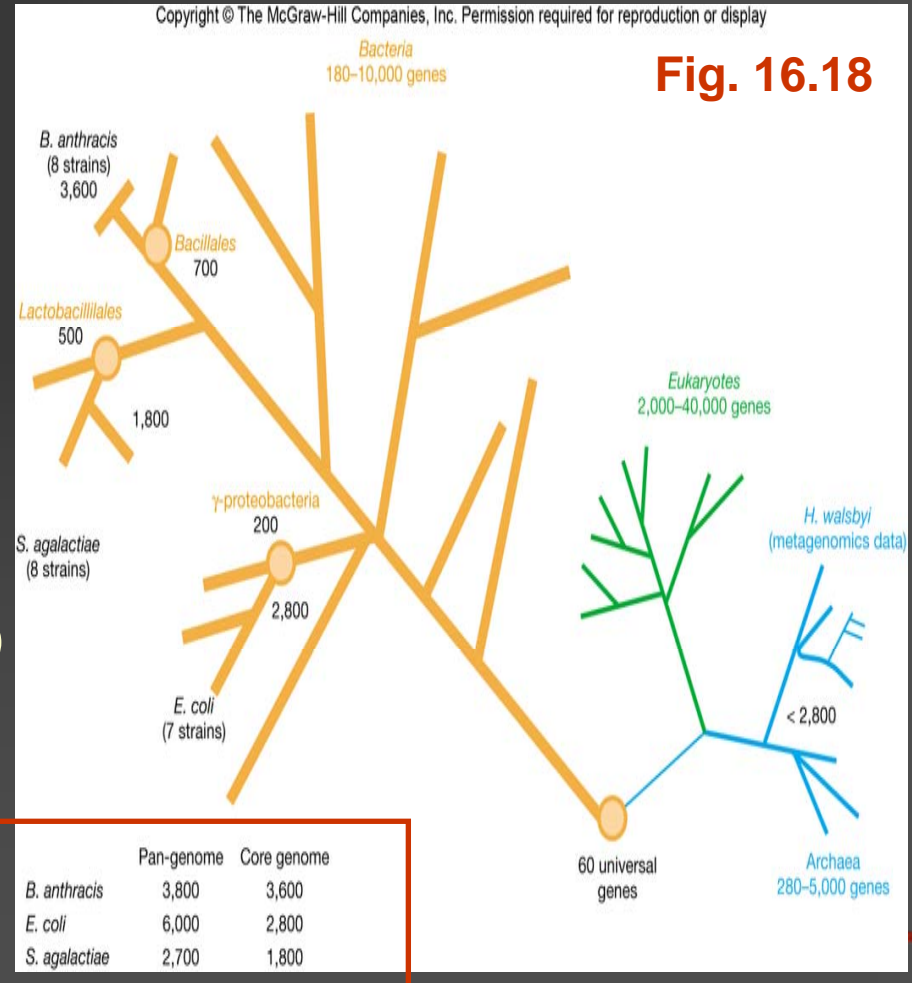
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# Core and Pan-genomes

- Core genome- set of genes found in all members of a species (or monophyletic group)
- Pan genome- combination of all different genes found in all the different strains in a given species
  - HGT (horizontal gene transfer)
  - Genomic island
  - Pathogenicity island
    - different GC contents
    - codon bias



# Synteny

- Phylogenetic relationship analysis
- To examine how similar the organization of orthologous genes are in the compared genomes

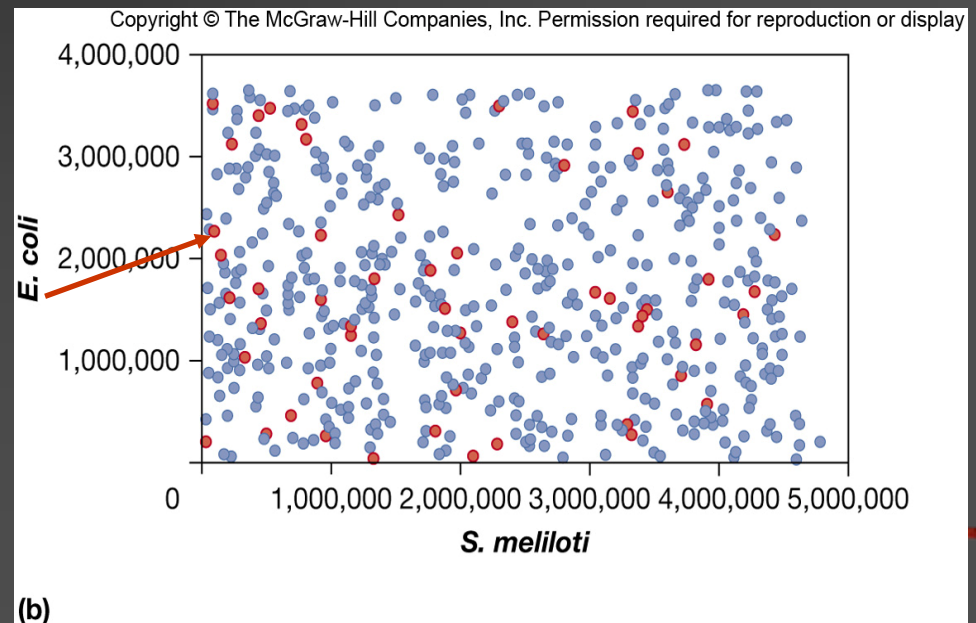
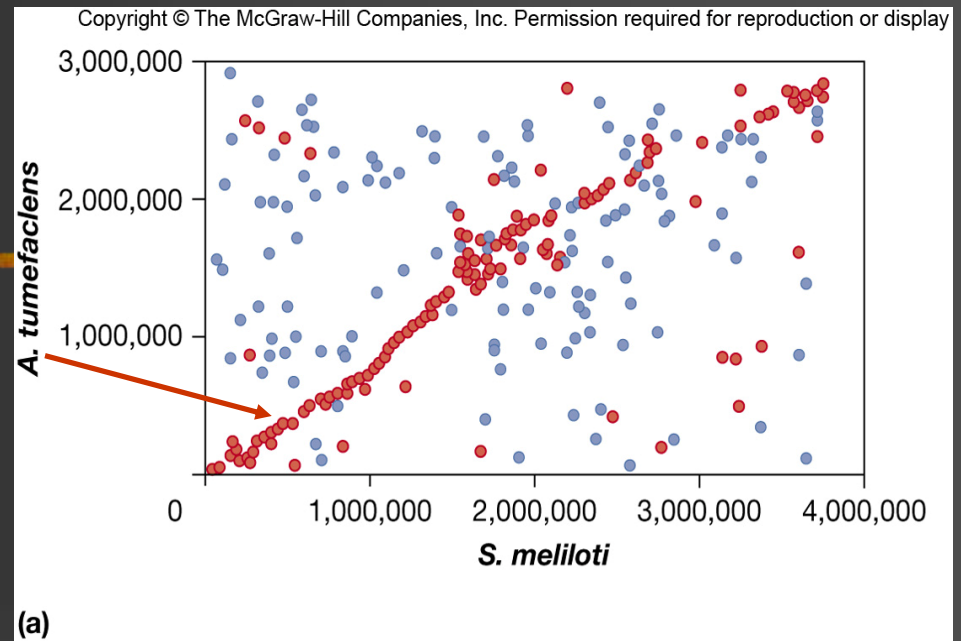
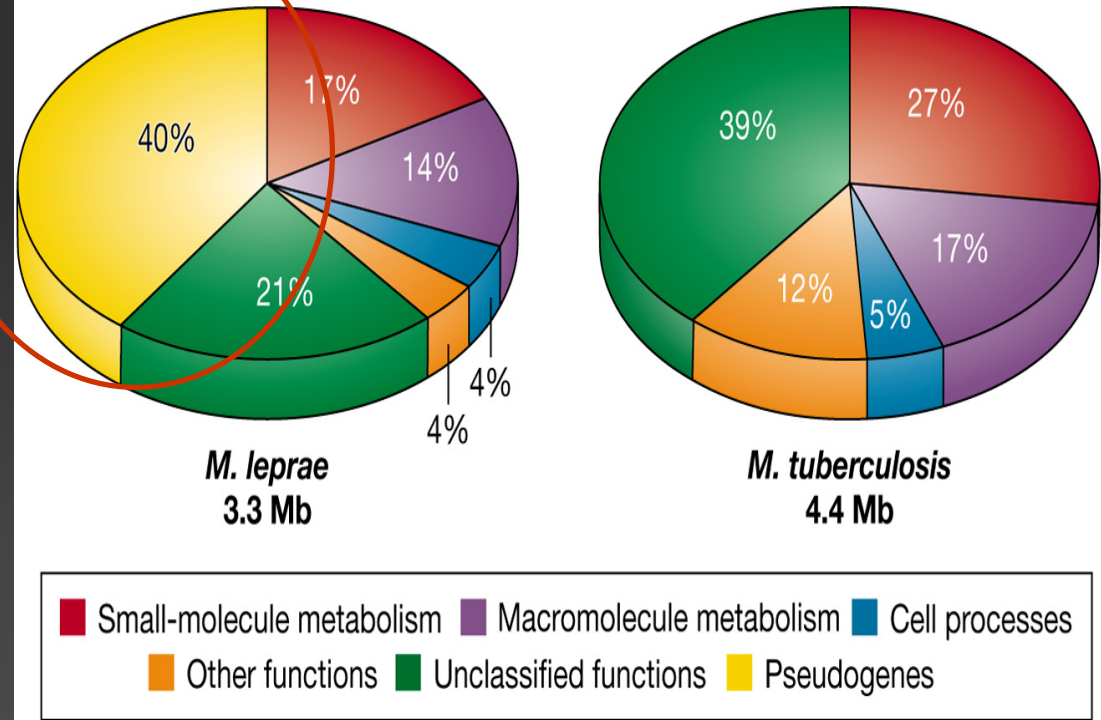


Fig. 16.19



# Comparative genomics analysis

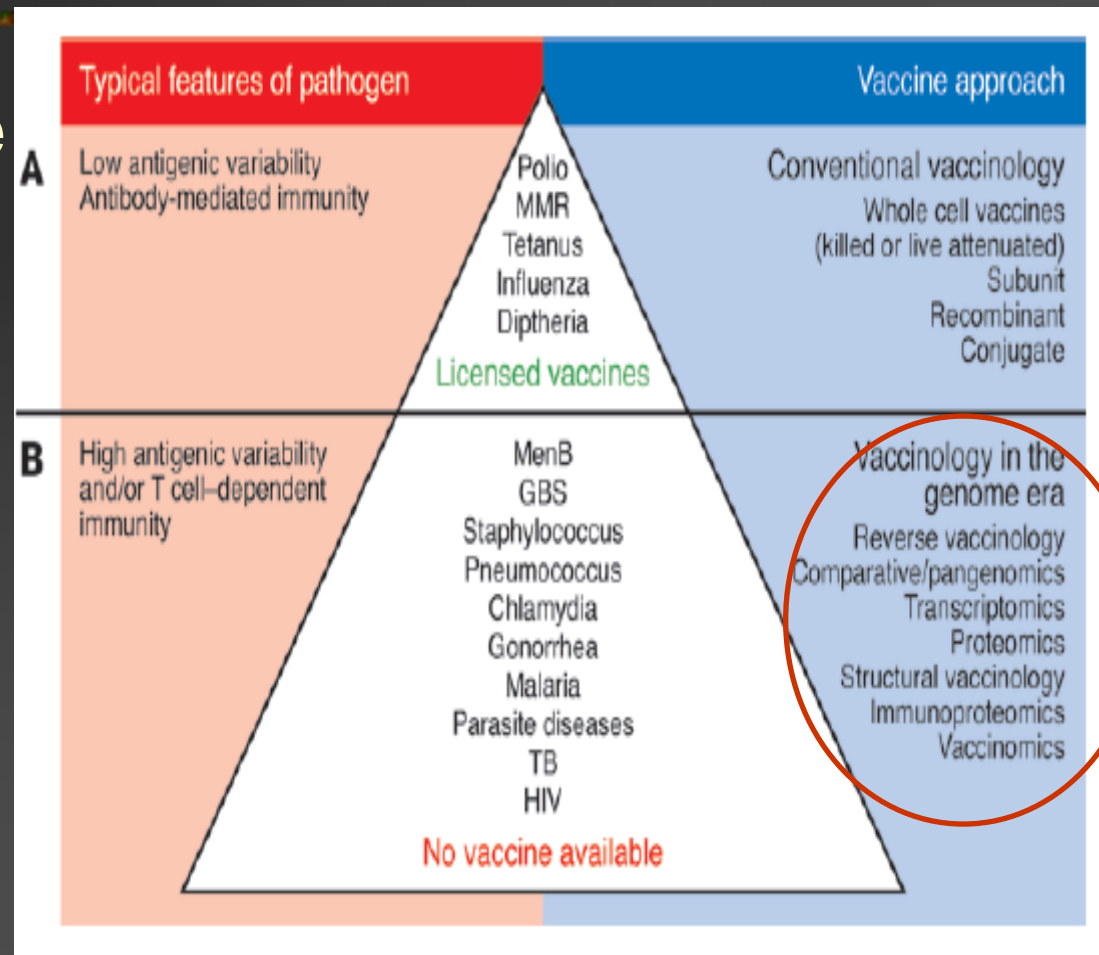
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- *M. leprae*- Leprosy
  - pseudogenes-degraded and nonfunctional genes
- *M. tuberculosis* (4.4 Mb) and *M. bovis* (4.3 Mb)- Tuberculosis
  - 99.5% sequence identity

# Reverse Vaccinology

- Two types of reverse vaccinology
  - Single species genome is examined for vaccine targets
  - a pan-genomic approach



# Metagenomics- an environmental genomics

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- used to learn more about the diversity and metabolic potential of microbial communities
    - shotgun sequencing DNA obtained directly from an environmental sample or series of related samples → determine the presence and level of classes of genes
    - may serve to establish hypotheses concerning interactions between community members
    - being viewed as a baseline technology for understanding the ecology and evolution of microbial ecosystems
-

# Construction and screening of genomic libraries directly from the environment

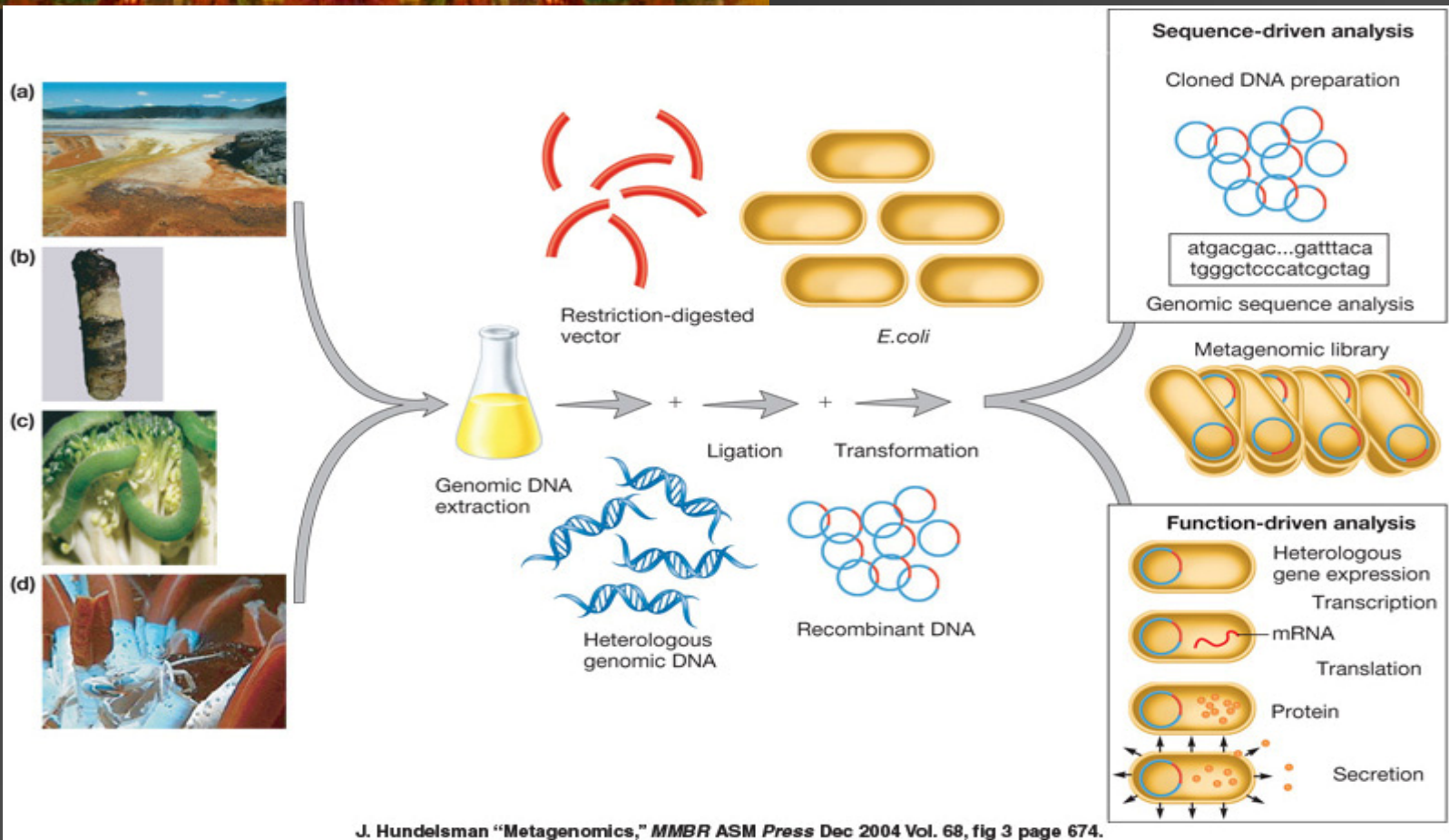
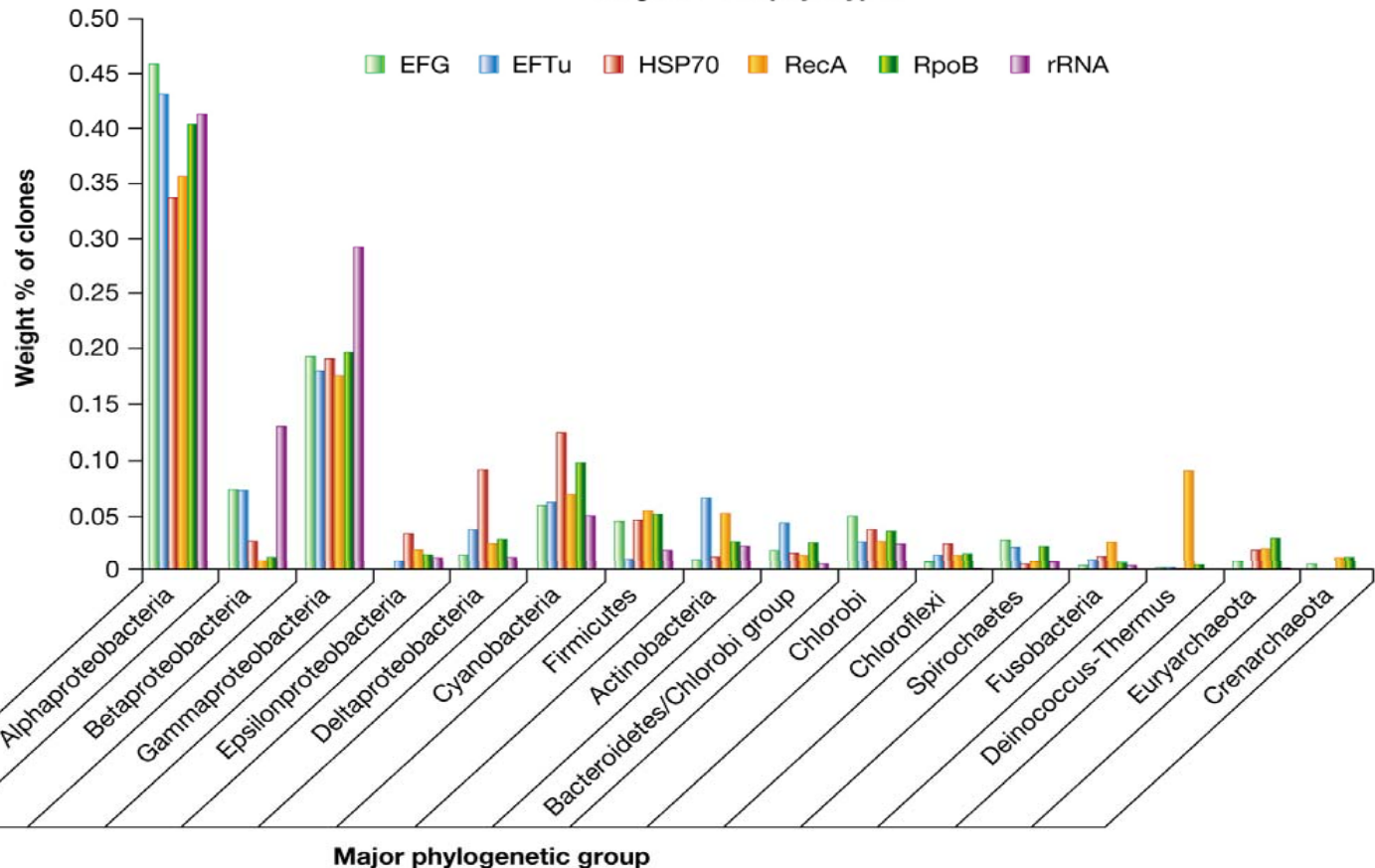


Figure 16.21

# Microbiome- phylogenetic diversity of Sargasso sea microbes

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Sargasso Sea phylotypes





# Sargasso sea

- portion of the Atlantic ocean surrounds Bermuda
  - The Sargasso Sea is well known for the floating mats of Sargassum weed





# The human microbiome project

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- Launched by the US National Institutes of Health (NIH) as one of its major roadmap initiatives (2007, earmarking ~US\$140 million)
  - to determine whether individuals share a core human microbiome
  - to understand whether changes in the human microbiome can be correlated with changes in human health
  - to develop the technological tools to support these goals
  - to address the ethical, legal and social implications of human microbiome research

Nature (2007) October 449:803-

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# Gut microbiome

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- The human intestine is home to ~100 trillion microorganisms of at least 400 species
  - $10^{11}$ ~ $10^{12}$  /ml bacteria in colon
  - Many gastrointestinal (GI) diseases are expected to be associated with disruption of host-bacterial interactions

PLOS PATHOGENS (2008)

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